



# **The biodiversity and biotechnological potential of marine fungi associated with sponges**

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# Sponges are relevant for secondary metabolites production

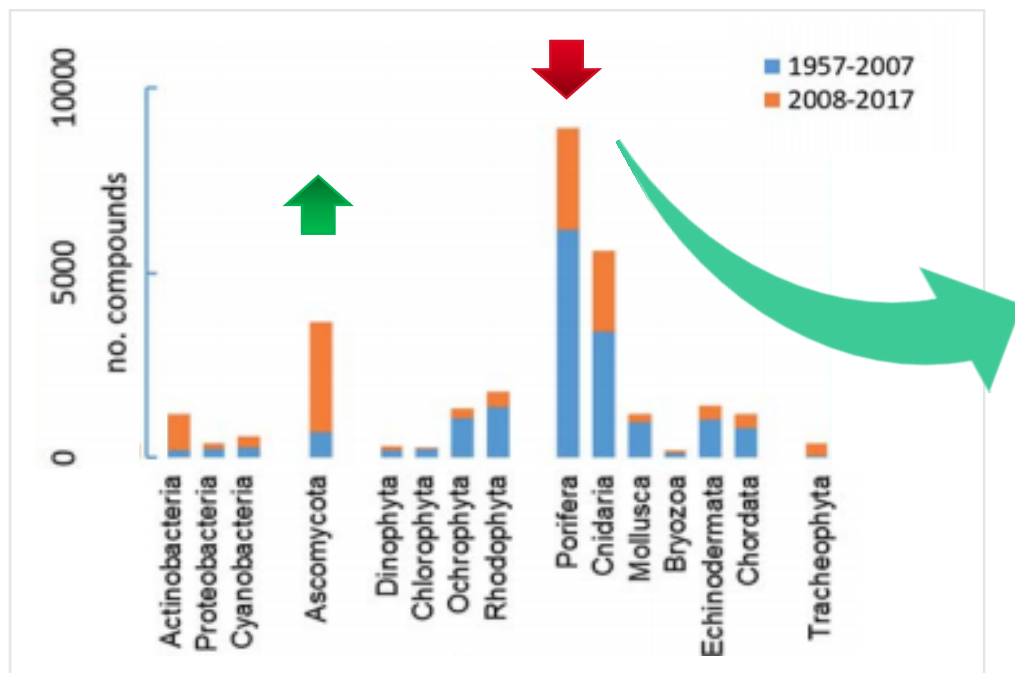
In 1951 the chemistry of marine natural products began with the isolation from *Cryptotethya crypta* of spongouridin and spongothymidin used as base structure for the new antiviral Ara-A (Anjum *et al.*, 2016)

## Ecological roles of secondary metabolites

Defense



- Predators
- Pathogens
- Fouling organisms



## Pharmacological properties

- Antibacterial
- Antifungal
- Anti-inflammatory
- Antiviral
- Anticancer
- Antituberculosis
- Anti-Alzheimer's
- Anti-HIV
- Antimalarial

## Environmental properties

- Antifouling

# Are sponges the real producer of secondary bioactive metabolites?

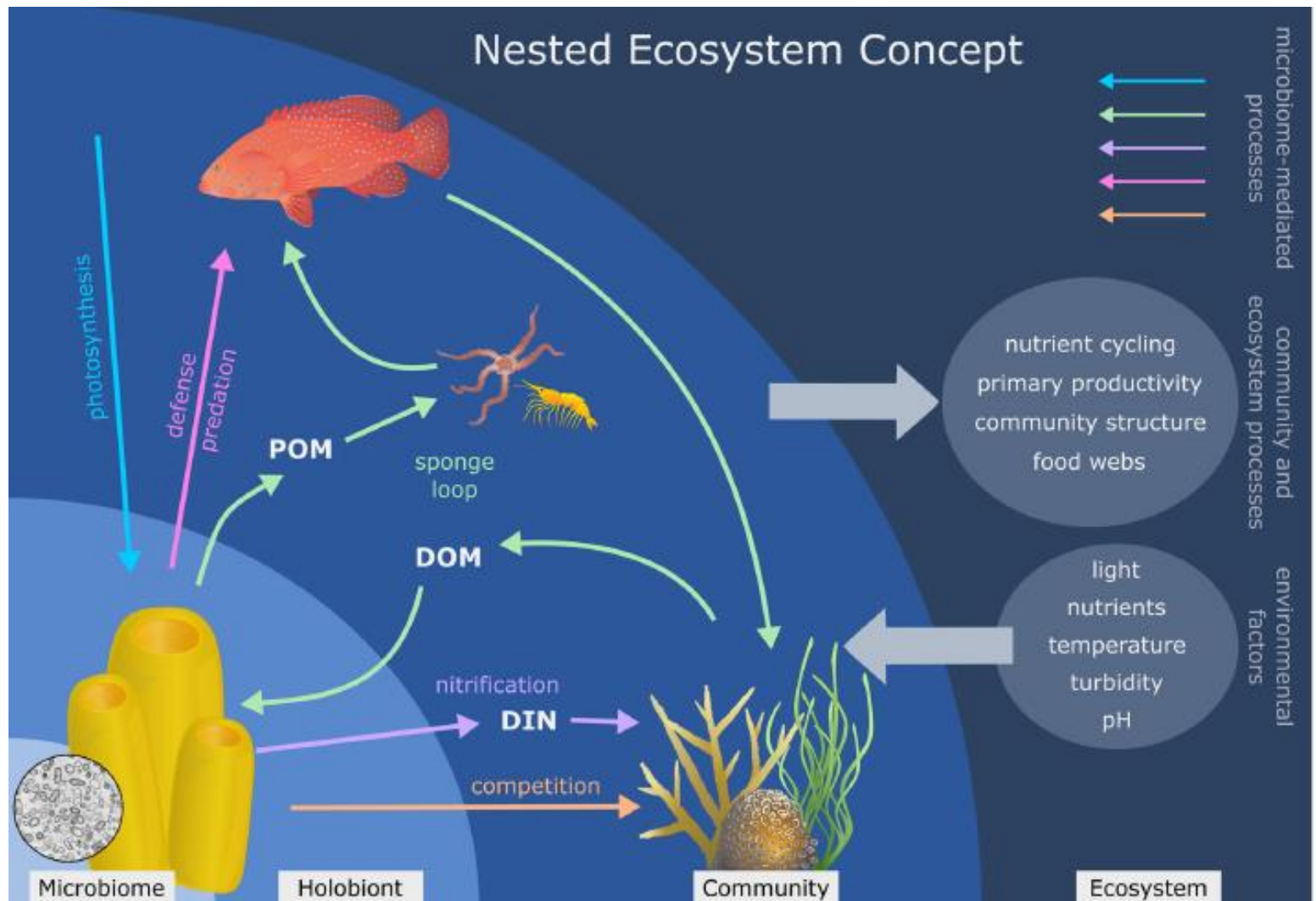
28% of novel compounds are produced by sponges associated fungi (Imhoff, 2016)

Over their 660–635 M years evolution sponges formed a close association with microorganisms including bacteria, archaea, fungi, and algae (Zumberge et al., 2018)

40-60% of sponges biomass is composed by microorganisms (Yarden, 2014)



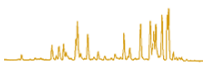
# The role of microorganisms within sponges: the “Sponge loop”



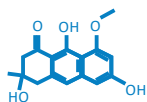




**Biodiversity of fungi associated with sponges**



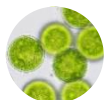
**Chemodiversity of fungi**



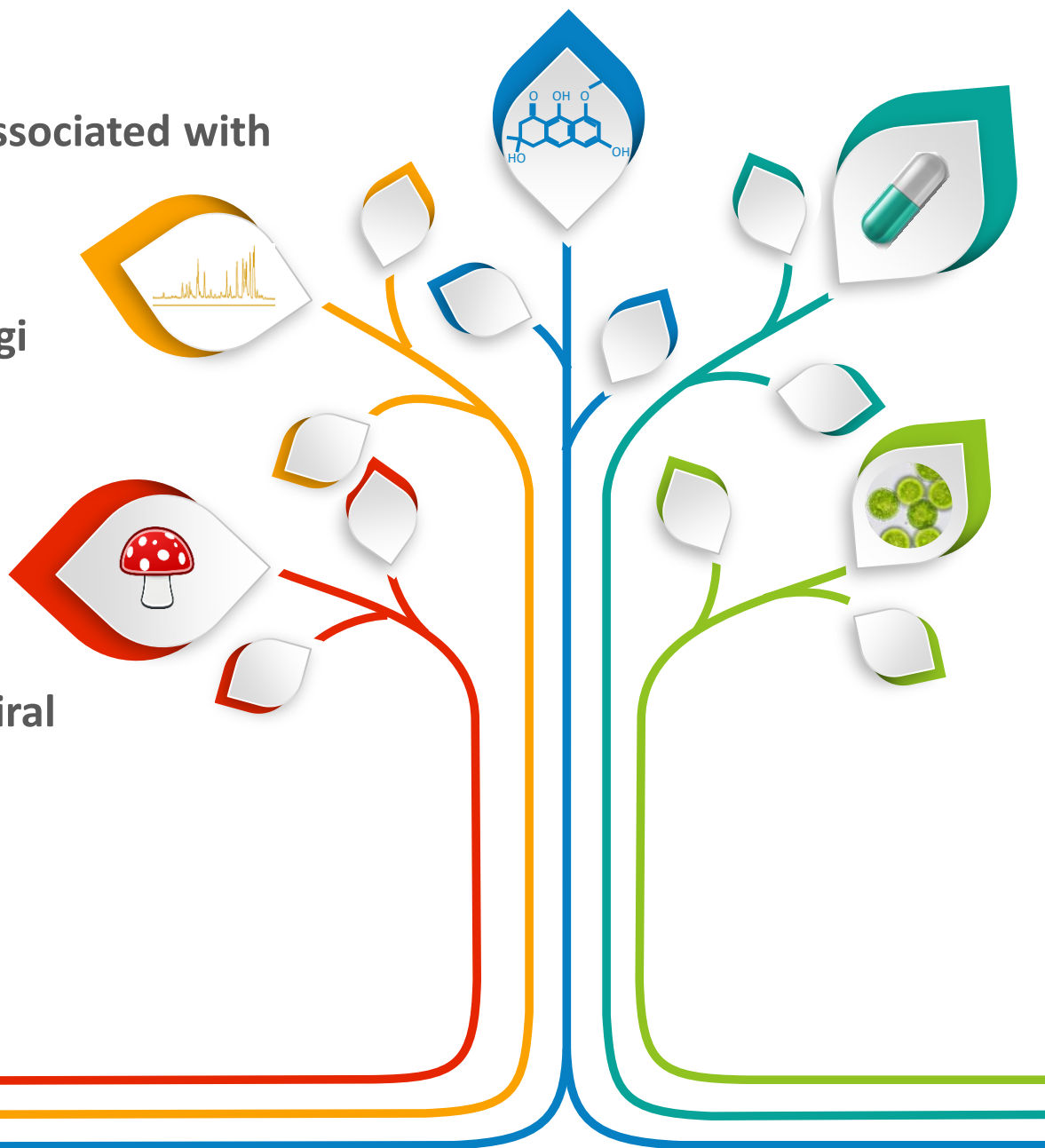
**Purification of fungal molecules**



**Antibacterial and antiviral activities**



**Antifouling activity**



# Four Atlantic species of sponges

Samples Collection



*Dysidea fragilis*  
Demospongiae



*Grantia compressa*  
Calcarea



*Pachymatisma johnstonia*  
Demospongiae

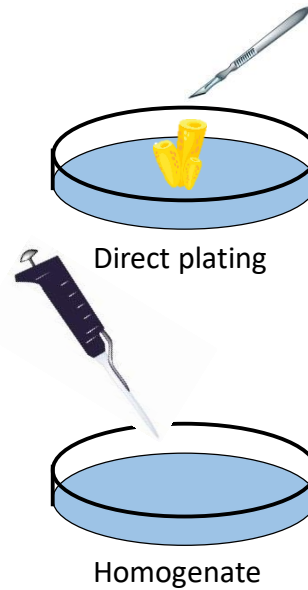
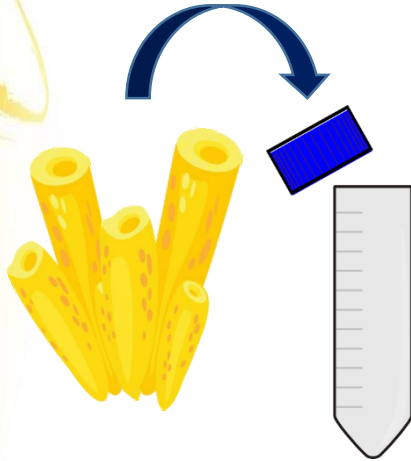


*Sycon ciliatum*  
Calcarea

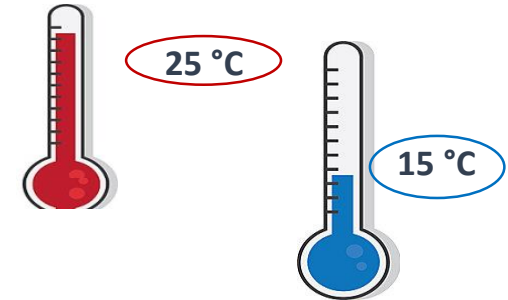


# Isolation and identification of fungi

Surface sterilization  
(ethanol 70%)



**4 Growth media:** SWA,  
CMA-SW, Gelatin-SW,  
MEASW

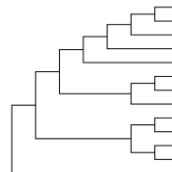
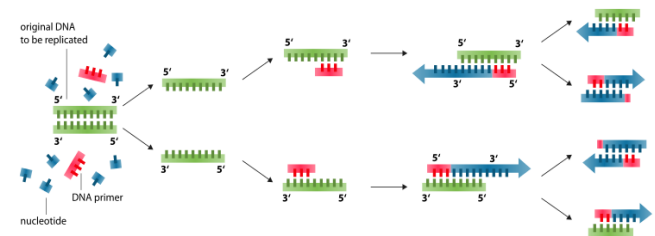


Pure cultures

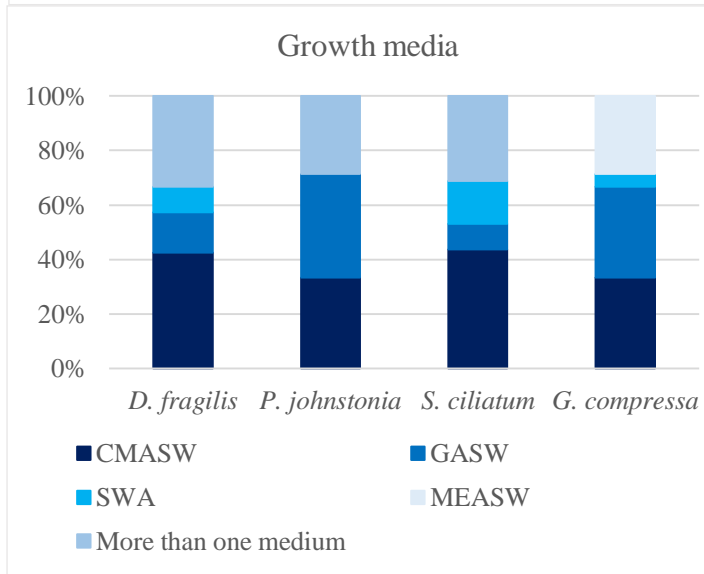
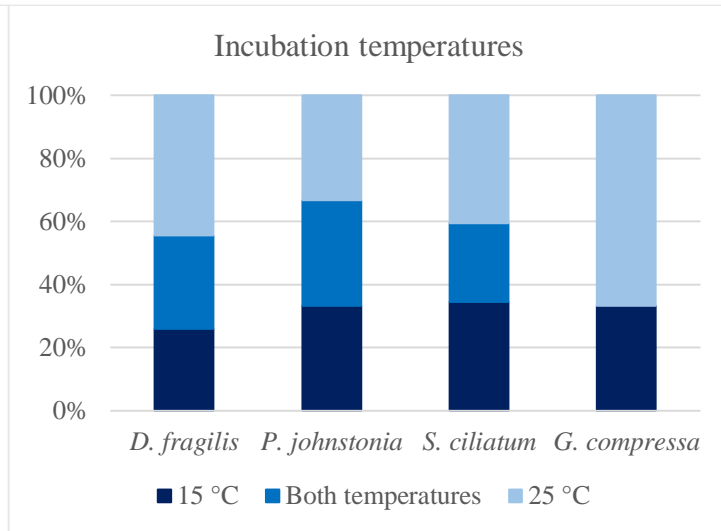
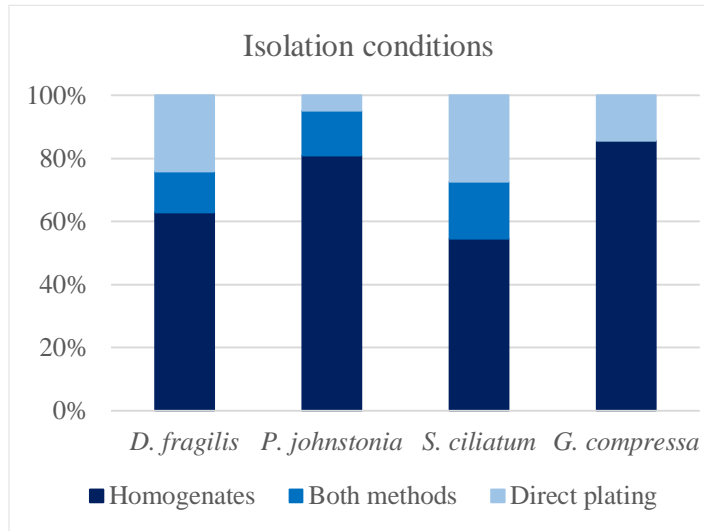
Morphological

Molecular

Phylogenetic



# Influence of the isolation techniques on fungal diversity



38% - 66% fungi were retrieved in only one of the conditions



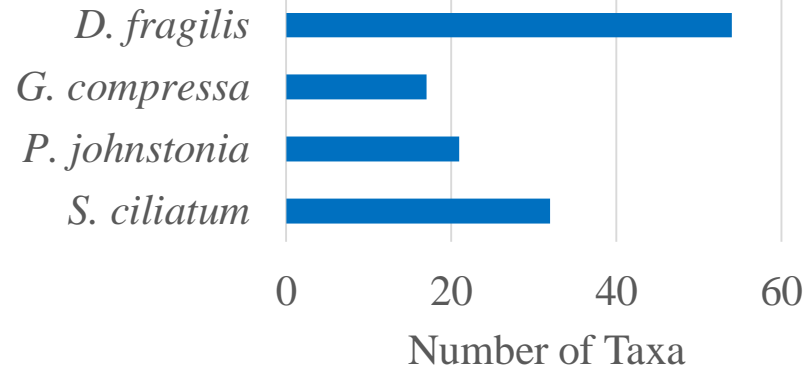
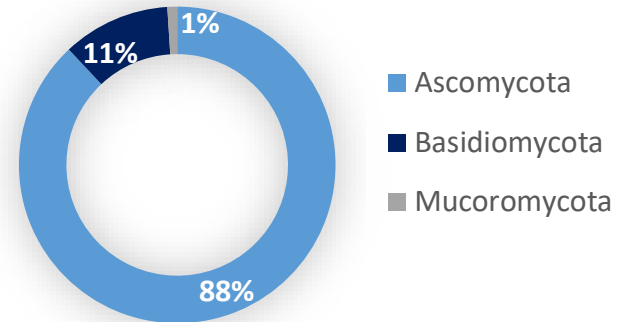
# Fungal diversity

97 taxa

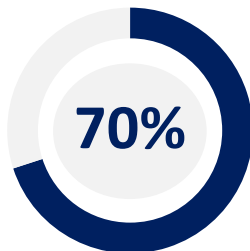


84% of taxa identified at species level

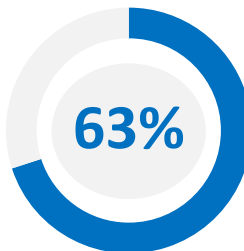
57 species → first report in sponges  
21 species → first report in marine environment



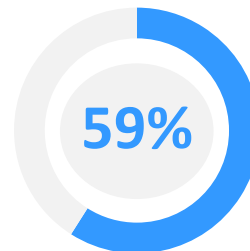
## Exclusive taxa



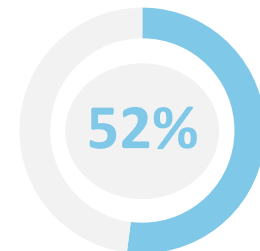
*D. fragilis*



*S. ciliatum*



*G. compressa*



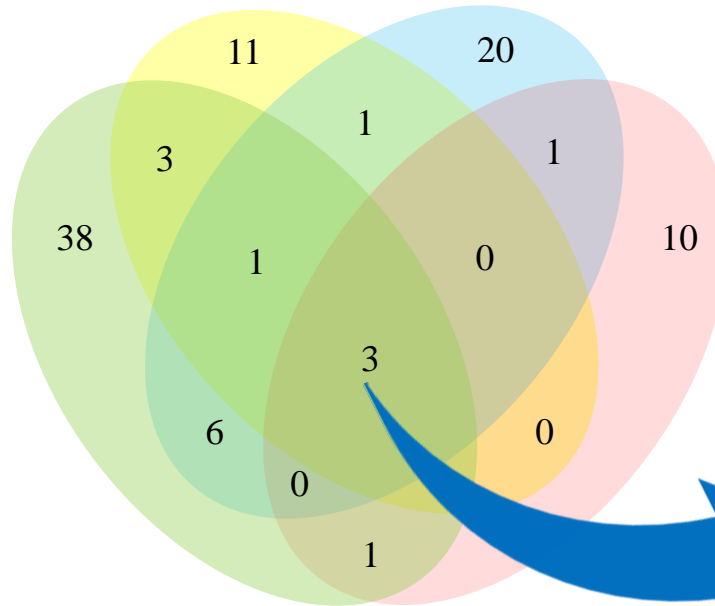
*P. johnstonia*

*P. johnstonia*

*S. ciliatum*

*D. fragilis*

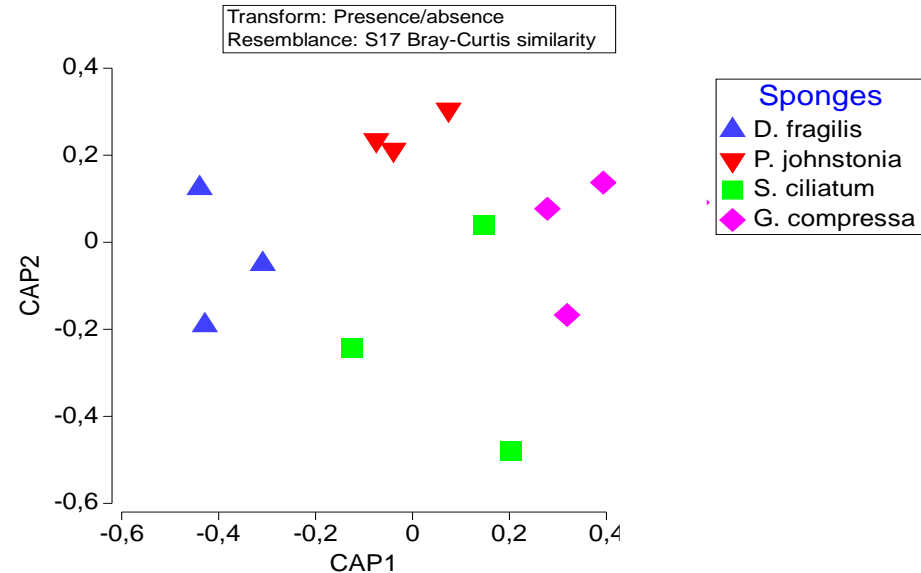
*G. compressa*



*Cladosporium cladosporioides*  
*Cladosporium allicinum*  
*Tolypocladium cylindrosporium*

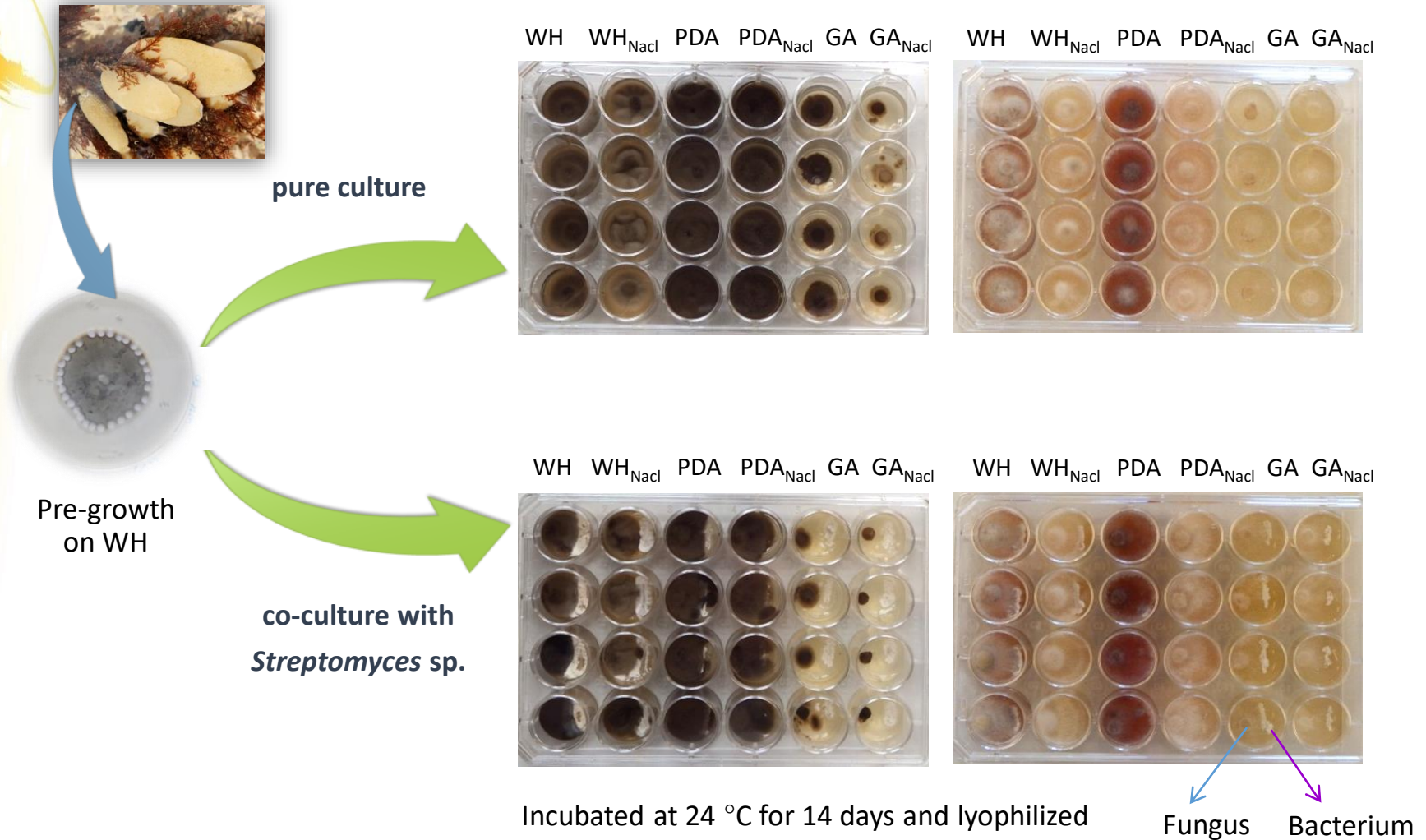
**The fungal community was significantly different ( $p < 0.01$ )**

Constrained Analysis of Principal Coordinates (CAP)

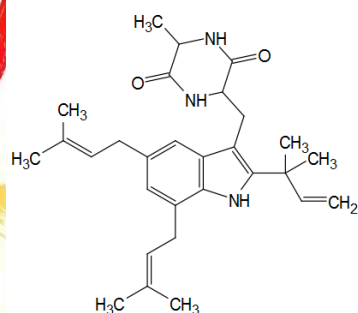


# OSMAC approach (One Strain – Many Compounds)

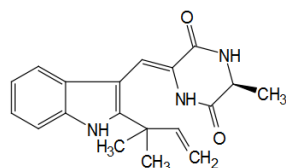
All 20 strains isolated from *G. compressa* were tested in 12 conditions



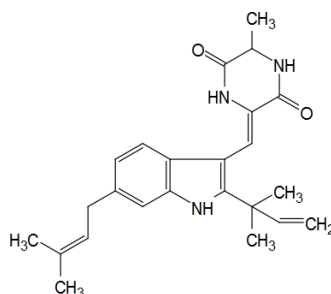
## 10 compounds isolated from *Eurotium chevalieri* MUT 2316



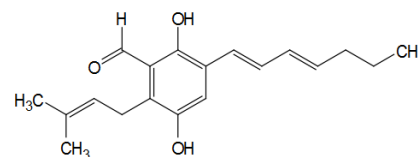
Echinulin  
(1) ●



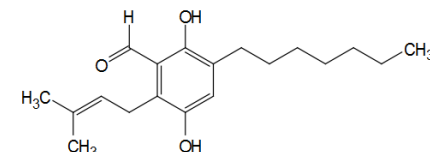
Neoechinulin A  
(2) ●



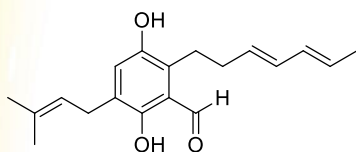
Neoechinulin D  
(3) ●



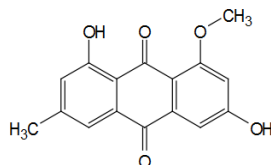
Dihydroauroglaucin  
(4) ●



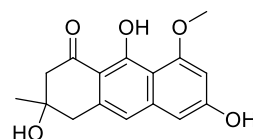
Flavoglaucin  
(5) ●



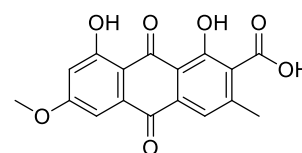
Isodihydroauroglaucin  
(6) ●



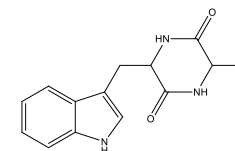
Phycion  
(7) ●



Asperflavin  
(8) ●



Cinnalutein  
(9) ●



Cyclo-L-Trp- L-Ala  
(10) ●

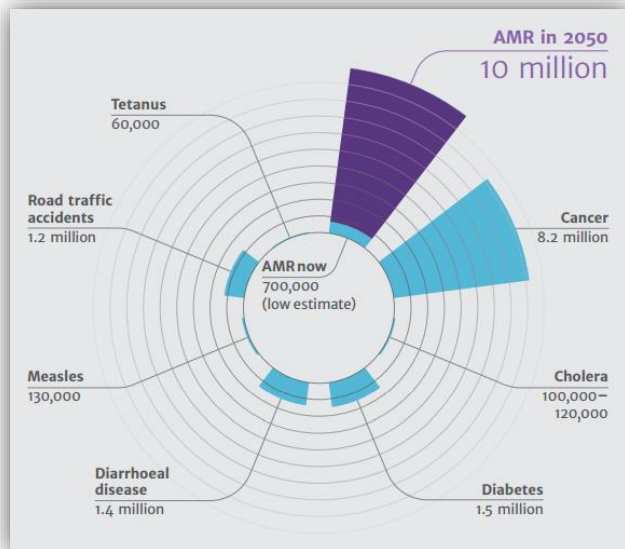
- Isolated for the first time from a marine fungus and in *E. chevalieri*
- Already recorded in *E. chevalieri* or its anamorph *Aspergillus chevalieri* isolated from soil, air or plants (Tawfik *et al.*, 2017; Zin *et al.*, 2017; Micheluz *et al.*, 2016; Kanokmedhakul *et al.*, 2011; Fraga *et al.*, 2008; Wu *et al.*, 2014; Ishikawa *et al.*, 1984; Hamasaki *et al.*, 1981)



# Which target for the 10 compounds produced by *E. chevalieri*?

## 1. Antibacterial activity

Antimicrobial resistance is now a serious threat to life quality and expectancy (Brown and Wright, 2016)



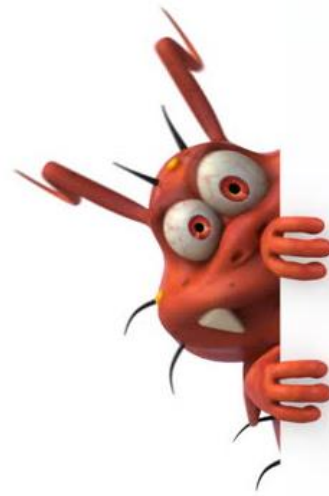
In 2050 multidrug resistant bacteria will be the first cause of death (O'Neill, 2016)

ANTIBIOTIC RESISTANCE IDENTIFIED		ANTIBIOTIC INTRODUCED	
Penicillin-R <i>Staphylococcus</i>	1940	1943	Penicillin
		1950	Tetracycline
		1953	Erythromycin
Tetracycline-R <i>Shigella</i>	1959	1960	Methicillin
Methicillin-R <i>Staphylococcus</i>	1962	1967	Gentamicin
Penicillin-R pneumococcus	1965	1972	Vancomycin
Erythromycin-R <i>Streptococcus</i>	1968		
		1979	Gentamicin-R <i>Enterococcus</i>
		1985	Imipenem and ceftazidime
Ceftazidime-R Enterobacteriaceae	1987		
Vancomycin-R <i>Enterococcus</i>	1988		
		1996	Levofloxacin
Levofloxacin-R pneumococcus	1996		
Imipenem-R Enterobacteriaceae	1998		
XDR tuberculosis	2000	2000	Linezolid
Linezolid-R <i>Staphylococcus</i>	2001		
Vancomycin-R <i>Staphylococcus</i>	2002		
PDR- <i>Acinetobacter</i> and <i>Pseudomonas</i>	2004/5	2003	Daptomycin
		2009	Ceftaroline
Ceftriaxone-R <i>Neisseria gonorrhoeae</i>	2009		
PDR-Enterobacteriaceae		2010	Ceftaroline
Ceftaroline-R <i>Staphylococcus</i>	2011		

From Ventola, 2015

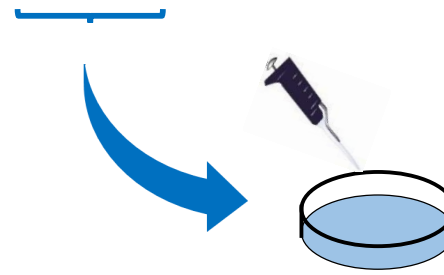
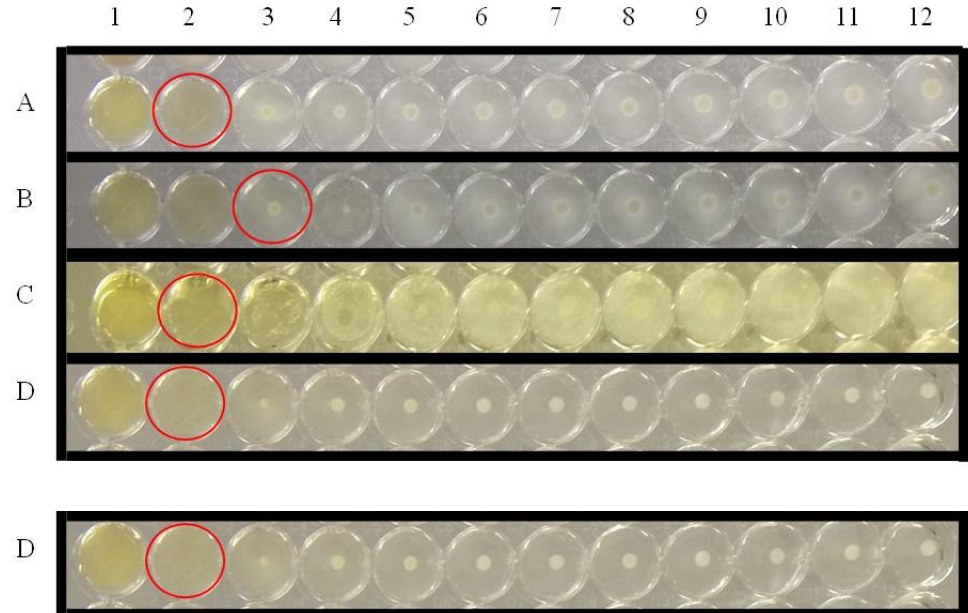
# Antibacterial activity

Species	Code	Gram +/-	Resistance
<i>Enterococcus faecalis</i>	ATCC 29212	+	-
<i>Escherichia coli</i>	ATCC 25922	-	-
<i>Pseudomonas aeruginosa</i>	ATCC 27853	-	-
<i>Staphylococcus aureus</i>	ATCC 29213	+	-
<i>Staphylococcus aureus</i>	Monza-PFI	+	Methicillin resistant
<i>Staphylococcus aureus</i>	Monza-FD1	+	Fluoroquinolone-resistant
<i>Streptococcus pneumoniae</i>	ATCC 49619	+	-
<i>Streptococcus pneumoniae</i>	Monza-82	+	Macrolide-resistant



**Minimal Inhibitory concentration (MIC  $\mu\text{g/mL}$ )**  
the lowest concentration of antimicrobial agent causing visible inhibition of bacterial growth

**Minimal bactericidal concentration (MBC  $\mu\text{g/mL}$ )**  
the lowest concentration of antimicrobial agent needed to kill bacteria



## The molecules were active against Gram + bacteria

Molecules	<i>S. aureus</i>		<i>S. aureus</i>		<i>S. aureus</i>		<i>E. faecalis</i>		<i>S. pneumoniae</i>		<i>S. pneumoniae</i>	
	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
Neoechinulin D (3) ★	-	-	-	-	-	-	-	-	-	-	64	-
Dihydroauroglaucin (4) ★	128	-	128	-	128	-	64	-	-	-	8	32
Isodihydroauroglaucin (6)	64	128	64	128	32	64	64	-	-	-	4	16
Physcion (7)	-	-	-	-	-	-	-	-	-	-	16	-
Asperflavin (8)	64	-	128	-	64	-	-	-	-	-	32	128
Cinnalutein (9) ★	-	-	-	-	128	-	-	-	-	-	32	-

"-" → >128 µg/mL

Methicillin resistant
  Fluoroquinolone resistant
  Macrolide resistant

MIC values lower than that reported for Gentamicin

MIC values lower than that reported for Erythromycin

★ First report of the antibacterial activity

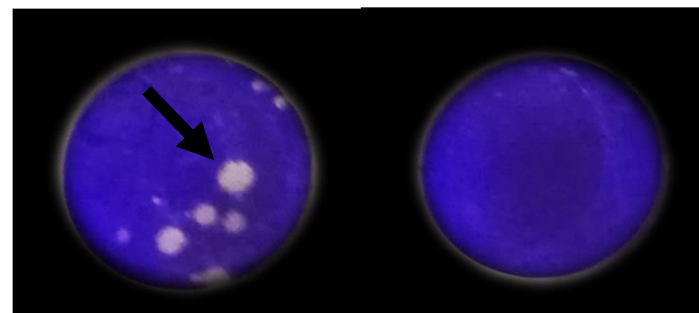


## 2. Antiviral activity

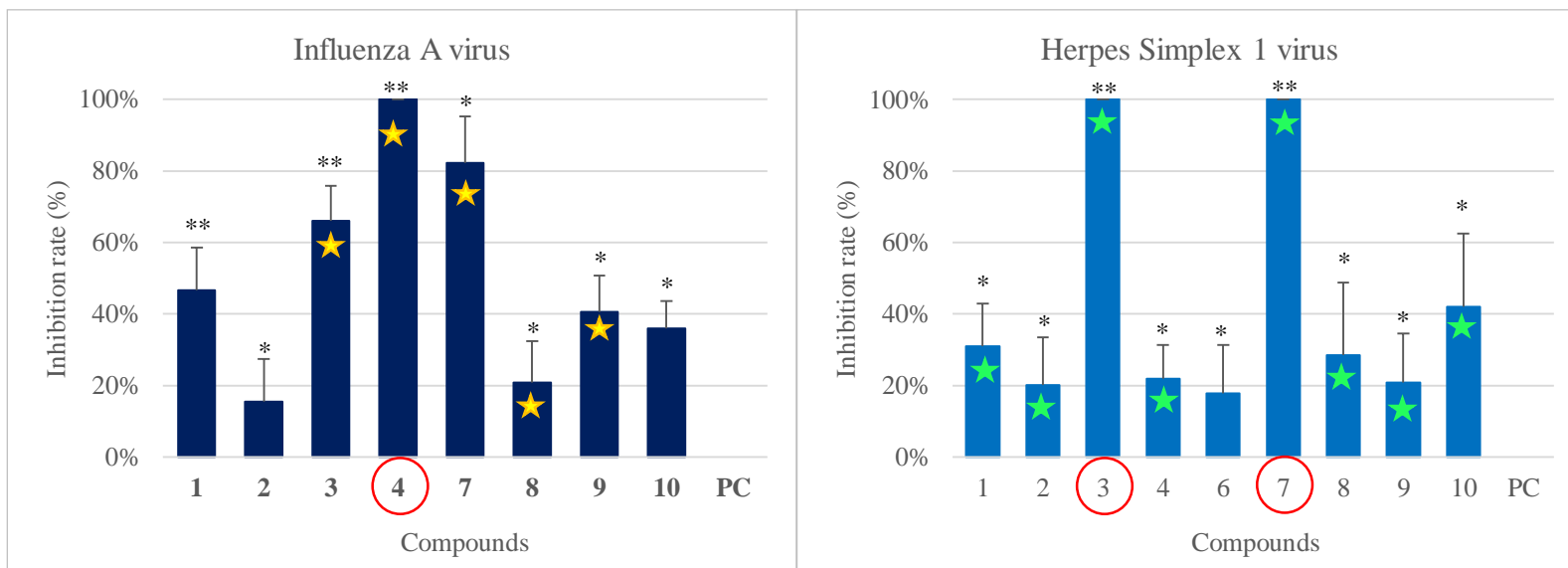
The antiviral activity was assessed against **Influenza A virus** and **Herpes Simplex Virus 1**

Positive control

100% inhibition



All the compounds inhibit the viral replication of Influenza A virus and Herpes Simplex 1 Virus



\*  $p < 0.05$  \*\*  $p < 0.01$

Dihydroauroglauцин (**4**) completely inhibited the Influenza A virus

Neoechinulin D (**3**) and Physcion (**7**) completely inhibited the Herpes Simplex 1 virus

★ First report of the antiviral activity against Influenza A virus

★ First report of the antiviral activity against Herpes Simplex 1 virus



### 3. Antifouling activity

**Marine biofouling: accumulation of microorganisms, algae and aquatic animals on surfaces immersed in seawater** (Amara *et al.*, 2018)

#### Consequences:

- Up to 40% Increased fuel consumption
- Increased carbon dioxide and sulphur dioxide emissions
- Invasive species might be transported by ships

#### Employed antifouling:

- Tributyltin based compounds → banned in 2008
- copper based paints → not environmental friendly

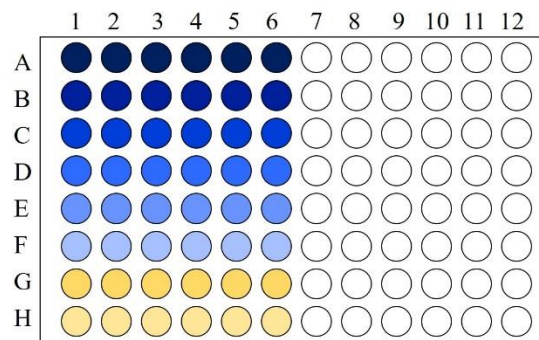


New and environmental friendly formulations are needed

Seven molecules were assessed for the ability to **inhibit the growth (Gr) and adhesion (Ad) of 5 marine bacteria and 5 algae representative of fouling organisms**

Bacteria	ATCC code
<i>Halomonas aquamarina</i>	14400
<i>Polaribacter irgensii</i>	700398
<i>Roseobacter litoralis</i>	49566
<i>Vibrio aestuarianus</i>	35048
<i>Pseudoalteromonas citrea</i>	29720

Microalgae	AC Code
<i>Cylindrotheca closterium</i>	170
<i>Exanthemachrysis gayraliae</i>	15
<i>Halamphora coffeaeformis</i>	713
<i>Porphyridium purpureum</i>	122
<i>Phaeodactylum tricornutum</i>	171



Compound 1  
A 1 to 6 (100 µg/mL)  
B 1 to 6 (10 µg/mL)  
C 1 to 6 (1 µg/mL)  
D 1 to 6 (0.1 µg/mL)  
E 1 to 6 (0.01 µg/mL)  
F 1 to 6 (0.001 µg/mL)

Compound 2  
A 7 to 12 (100 µg/mL)  
B 7 to 12 (10 µg/mL)  
C 7 to 12 (1 µg/mL)  
D 7 to 12 (0.1 µg/mL)  
E 7 to 12 (0.01 µg/mL)  
F 7 to 12 (0.001 µg/mL)

G 1 to 12 Positive control  
H 1 to 12 Negative control

Growth and adhesion were monitored spectroscopically

Bioassays against **macrofoulers**



Inhibition of blue mussel  
*Mytilus edulis* settlement  
**tyrosinase assay**



**Low Observable Effect Concentration (LOEC):** the lowest concentration with an average response that is significantly different from the control

# Conclusions

Increase the knowledge of marine fungi associated with sponges. **97 taxa** were isolated

The **10 molecules** produced by *E. chevalieri* demonstrated an incredible **biotechnological potential**

**Neoechinulin D** and **Physcion** completely inhibited the **replication of Herpes Simplex 1 virus**. **Dihydroauroglaucin** completely inhibited the **replication of Influenza A virus**

Significant **difference** among the **mycobiotas of the sponges**

More than half of the **compounds were active against at least one bacterium**, with several molecules more efficient than already known antibiotics

**Cyclo-L-Trp- L-Ala** performed better in the antifouling tests showing **inhibition of tyronisase, antibacterial and antimicroalgal activities**

1

2

3

4

5

6



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Vinci Program and Galileo Program





Thanks for the attention