

Understanding the emergence of Group B Streptococcus neonatal infections by the genomic analysis of today's and collection isolates

Philippe Glaser

S. agalactiae or group B streptococcus (GBS)

- Opportunistic pathogen
- Human
 - Commensal of the digestive or urinary tract
 - Leading cause of neonatal infections
 - Emerged during the 1960s–1970s
 - Risk for immuno-compromised adults and elderly



- Broad host spectrum in animals
 - Udder infections in bovines and camels
 - Invasive diseases in fish - outbreaks

The emergence of GBS neonatal infections

Dtsch med Wochenschr 1965; 90(6): 258-261

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Streptococcus agalactiae als Erreger von Säuglingsmeningitiden

Streptococcus agalactiae as a cause of meningitis in the newborn (report of three cases)

G. Kexel, S. Schönbohm

Summary

Within **half a year three** independent cases of **meningitis** caused by *Streptococcus agalactiae* were observed (at the Children's Clinic, University of Bonn). The three infants were **under the age of three months** and the disease took a fulminating course leading to **death within hours**. The clinical picture of the three cases was very similar and was different from that observed in other types of meningitis.

Bacteriological differentiation of the causative organism is discussed. **The manner of infection remains unexplained.**

MLST versus Genome sequencing

MLST

- Based on the sequences of 7 genes
- 751 Sequence Types (ST)
- Performed by 57 laboratories from 29 countries
- No redundancy
- Easy to analyse but low resolution

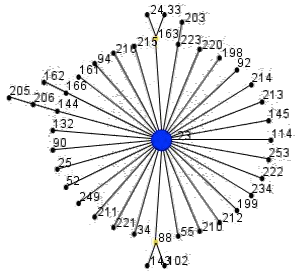
Genome sequencing

- Complete genomic information
- 506 available genome sequences
- Performed by few laboratories
- Specific sampling to respond specific questions

Population structure of GBS derived from MLST

CC23

Human
bovine



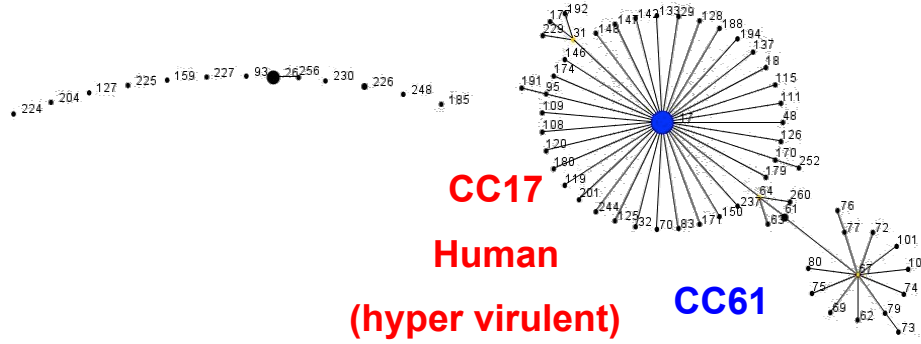
CC17

Human

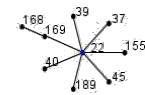
(hyper virulent)

CC61

bovine



CC22

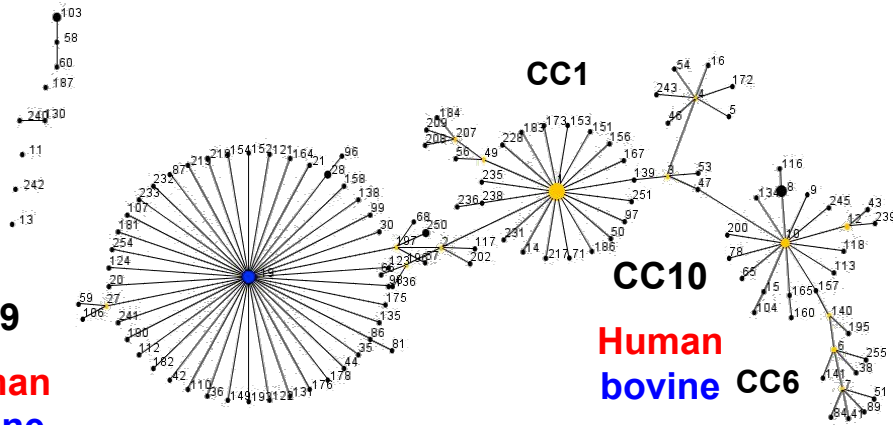


CC1

CC10

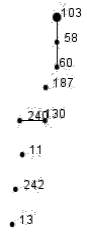
Human
bovine

CC6



CC19

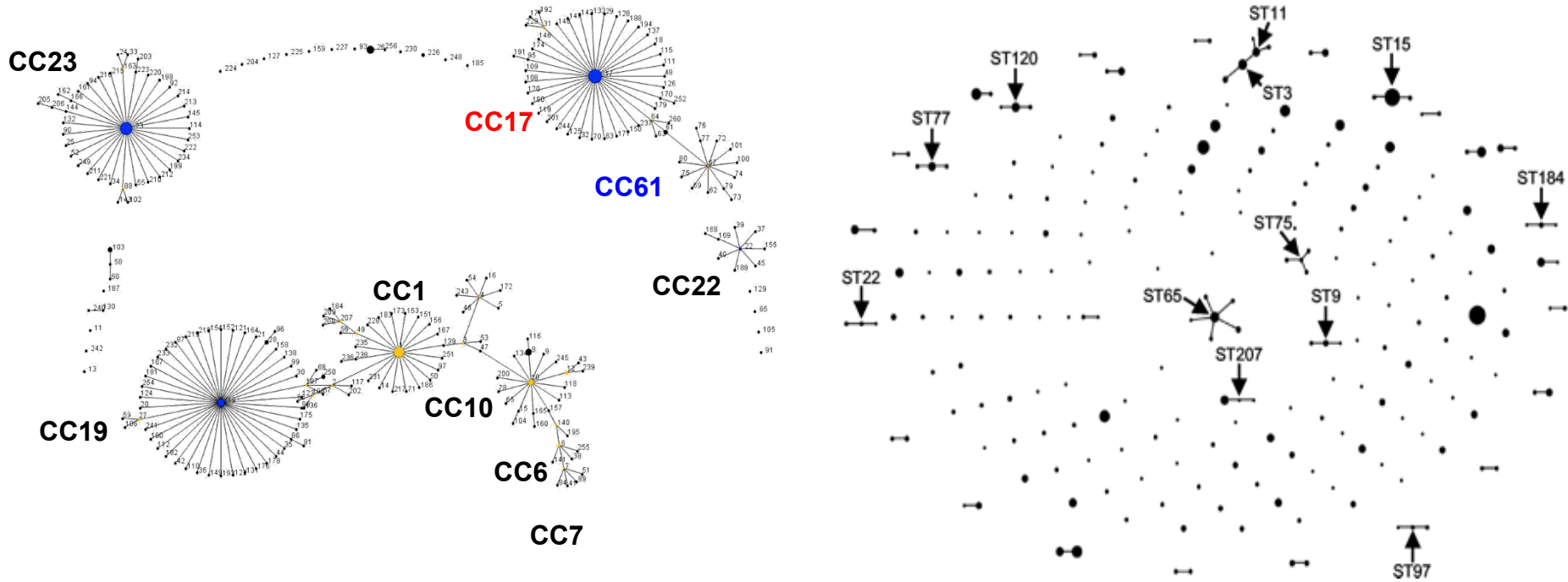
Human
bovine



CC7
Bovine
Human
Fish



GBS versus GAS population structures



⇒ Low complexity of GBS population

Population studies of GBS

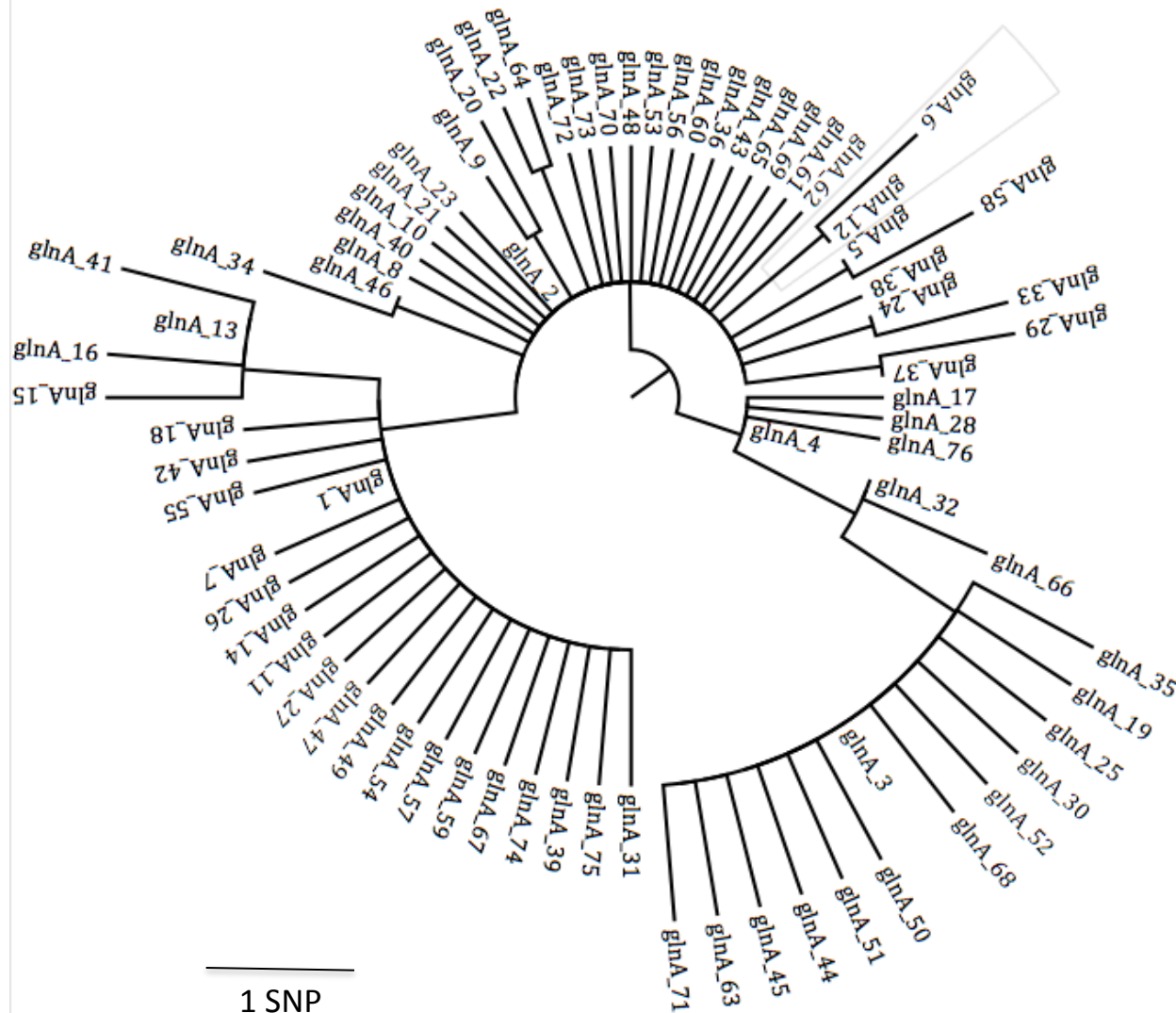
Study	Origin	Nb of isolates	CC1 (ST1)	CC8-10	CC17	CC19 (ST19, 28)	CC23	CC26	other
Jones	World	152	16% 14%	18%	30%	17% 13 - 2%	12%	2%	4%
Luan	Sweden	158	15% 9%	13%	24%	29% 20%	14%	0	4%
Manning	USA	338	29% 25%	18%	8%	17% 14%	25%	0,3%	2%
Bohnsack	Canada	899	16% 10%	9%	10%	17% 12%	40%	0	4%
Sadowi	Poland	112	18% 13%	18%	14%	13% 9 - 2%	37%	0	0
Huber	Kenya	169	12% 9%	17%	21%	13% 5%	27%	2%	7
Brochet	Dakar Bangui	163	20% 9%	6%	12%	28% 4% - 15%	17%	15%	2

New STs and clonal complexes

glnA ML tree
75 alleles

All recent alleles show single SNPs with older alleles

- Most of the phylogenetic diversity of the species is known
- New combination of STs



Distribution of genome sequences in STs

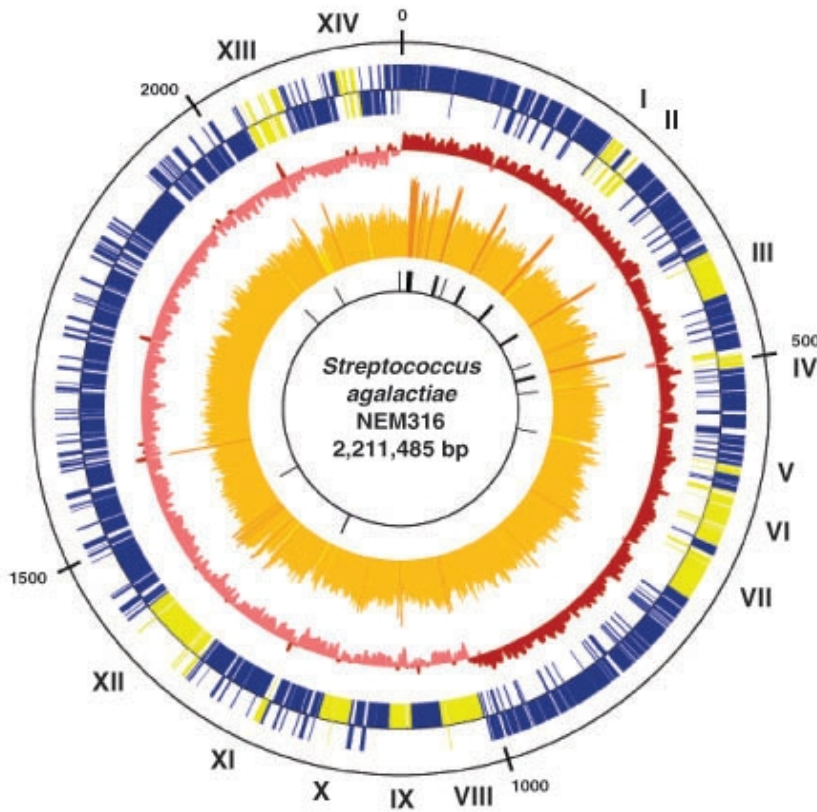


Belong to 59 STs

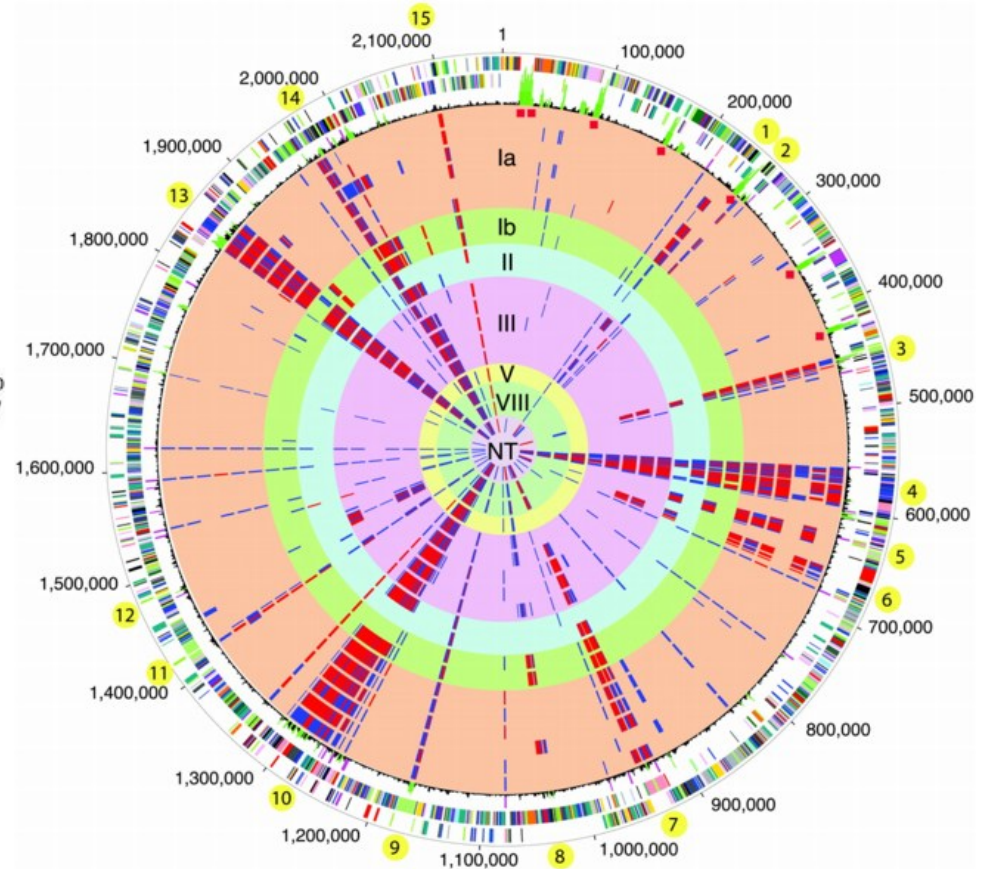
ST1:	78
ST7,8,10, 12:	47
ST17:	60
ST19:	53
ST23:	51
Unidentified:	93

- Complete genome sequencing covers only partially the diversity of the species
- Sequencing of specific isolates to fill gaps in the diversity

Sequencing 1 genome: a mosaic organization



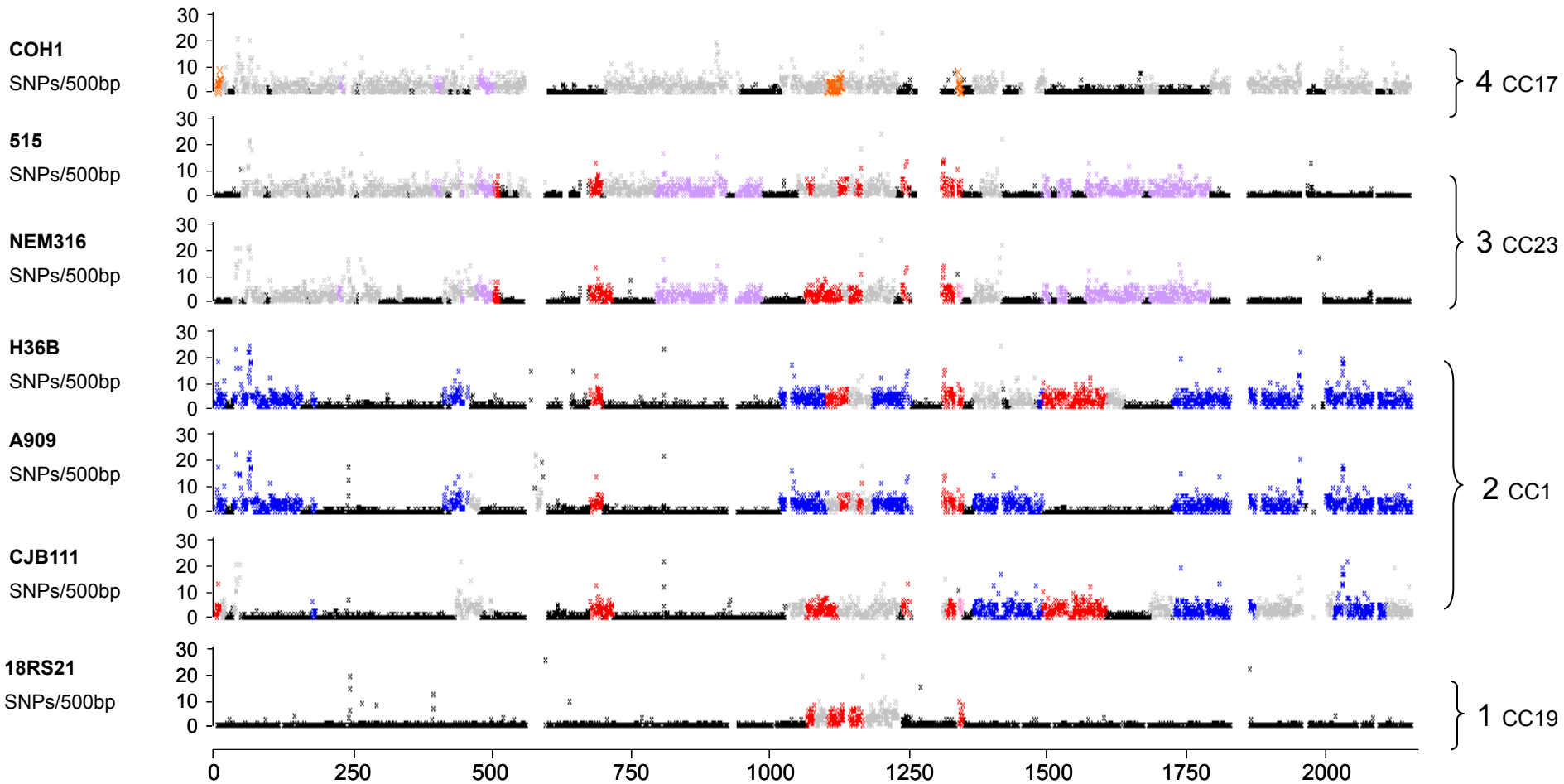
Glaser *et al.* 2002



Tettelin *et al.* 2002

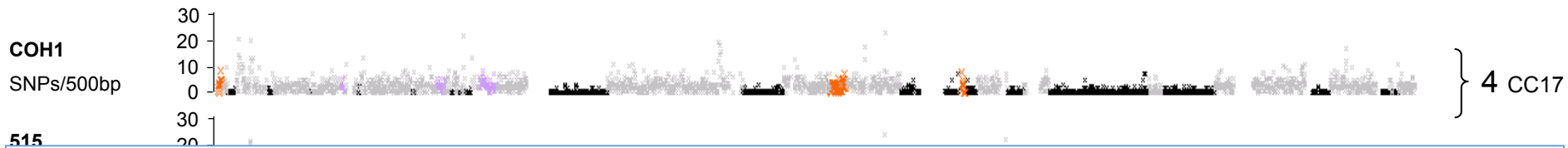
Polymorphisms among the 8 genome sequences

■ Identical to 2603 V/R ■ Specific to the strain ■ or ■ or ■ Region shared between 2 strains, at least

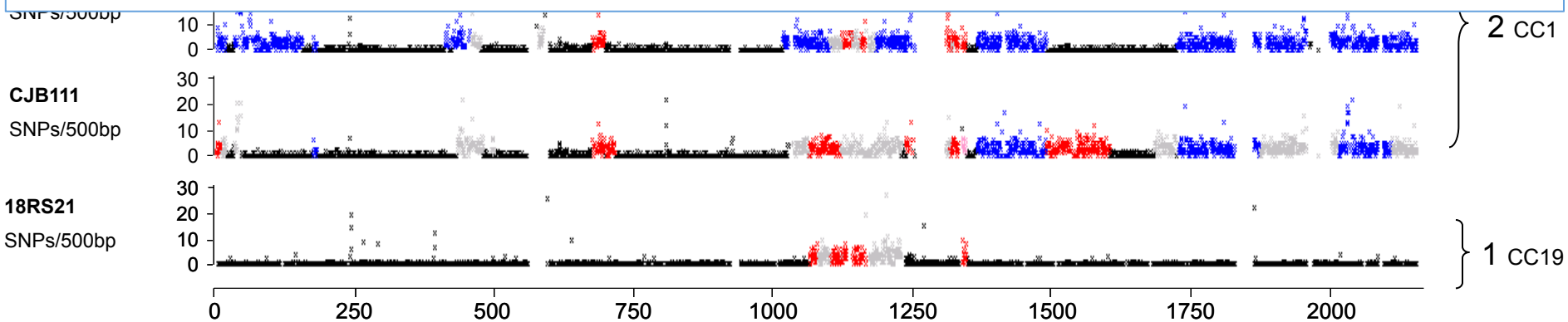


Polymorphisms among the 8 genome sequences

■ Identical to 2603 V/R ■ Specific to the strain ■ or ■ or ■ Region shared between 2 strains, at least



- *S. agalactiae* genomes are shaped by conjugative exchanges of large DNA regions
- Integrative and conjugative elements are responsible for these exchanges



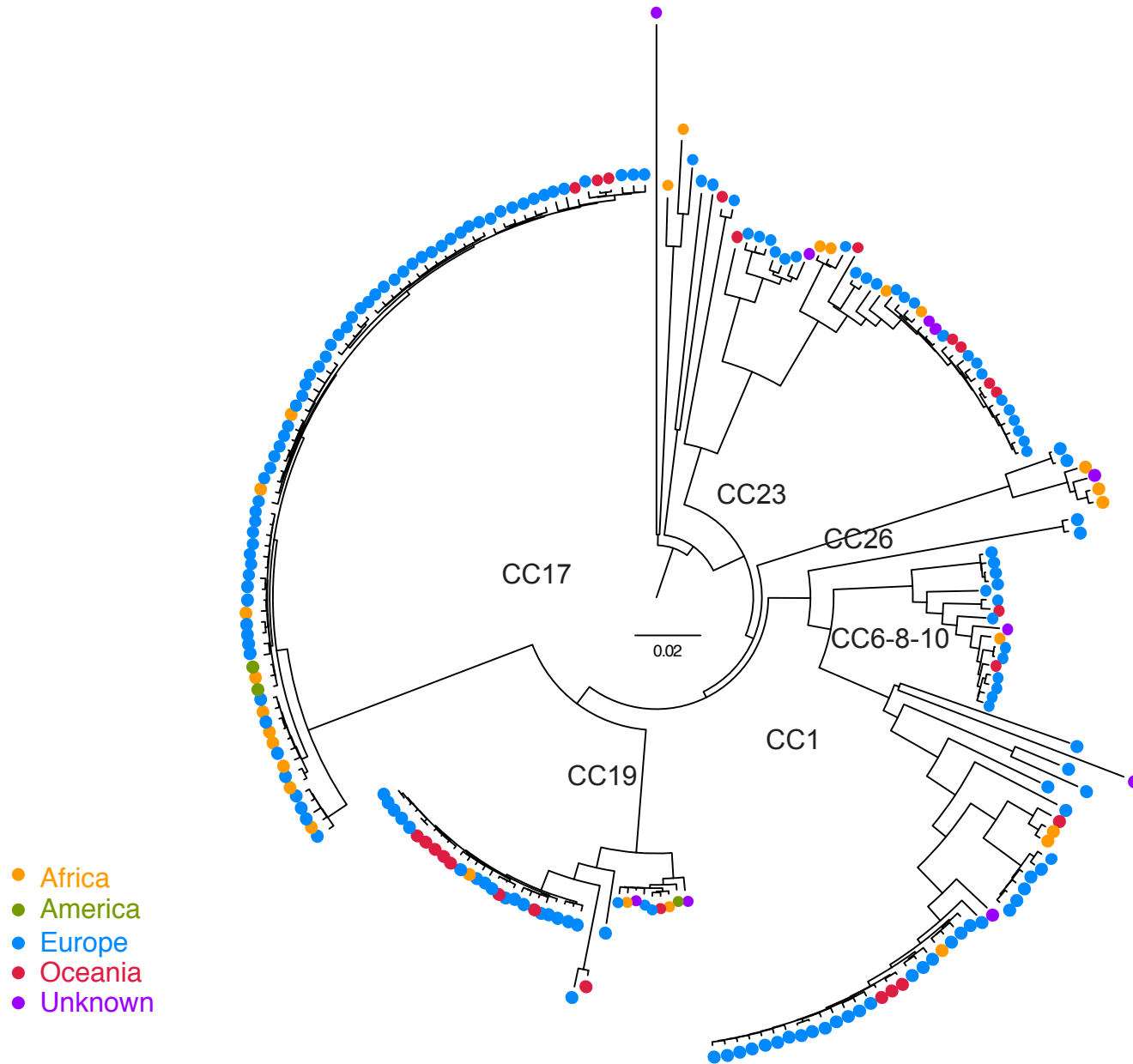
Population genomics of *S. agalactiae*

- Objective
 - To confirm the observed emergence of GBS infections at the bacterial population level
 - To get clues on the reasons for this emergence
- Strains:
 - Sanger Centre:
 - 92 European strains – DEVANI consortium (7 countries)
 - 24 Australian strains
 - Institut Pasteur:
 - 112 isolates: 27 from Africa, carriage and clinical isolates, 9 bovine strains.
 - 12 strains isolated between 1953 and 1961

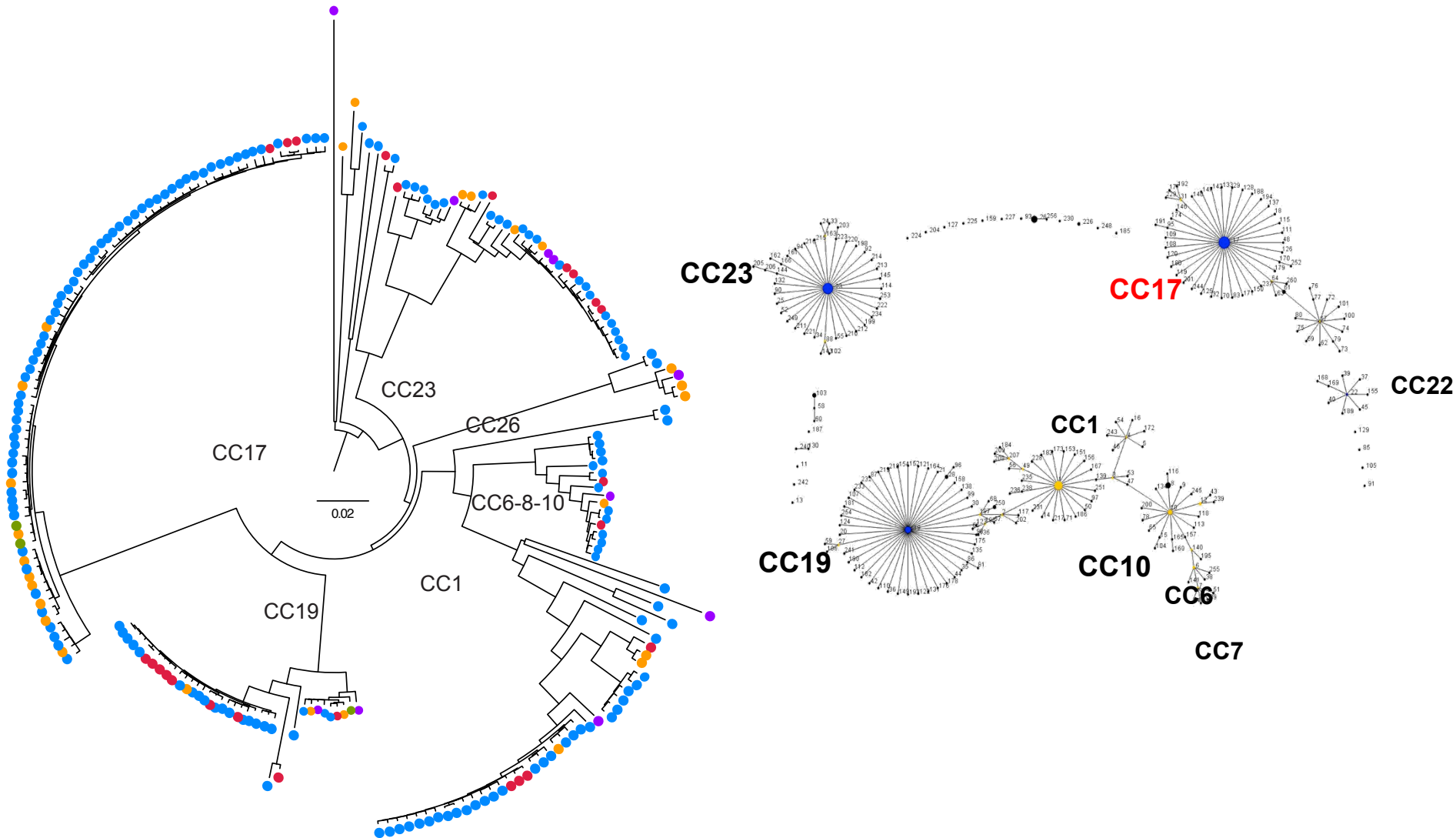
Methods

- Isolates were sequenced by the Illumina technology
- Assembled by using Velvet
- SNP were identified by using BWA and Mpileup (Samtools)
- Phylogeny was performed by maximum-likelihood and by using BEAST

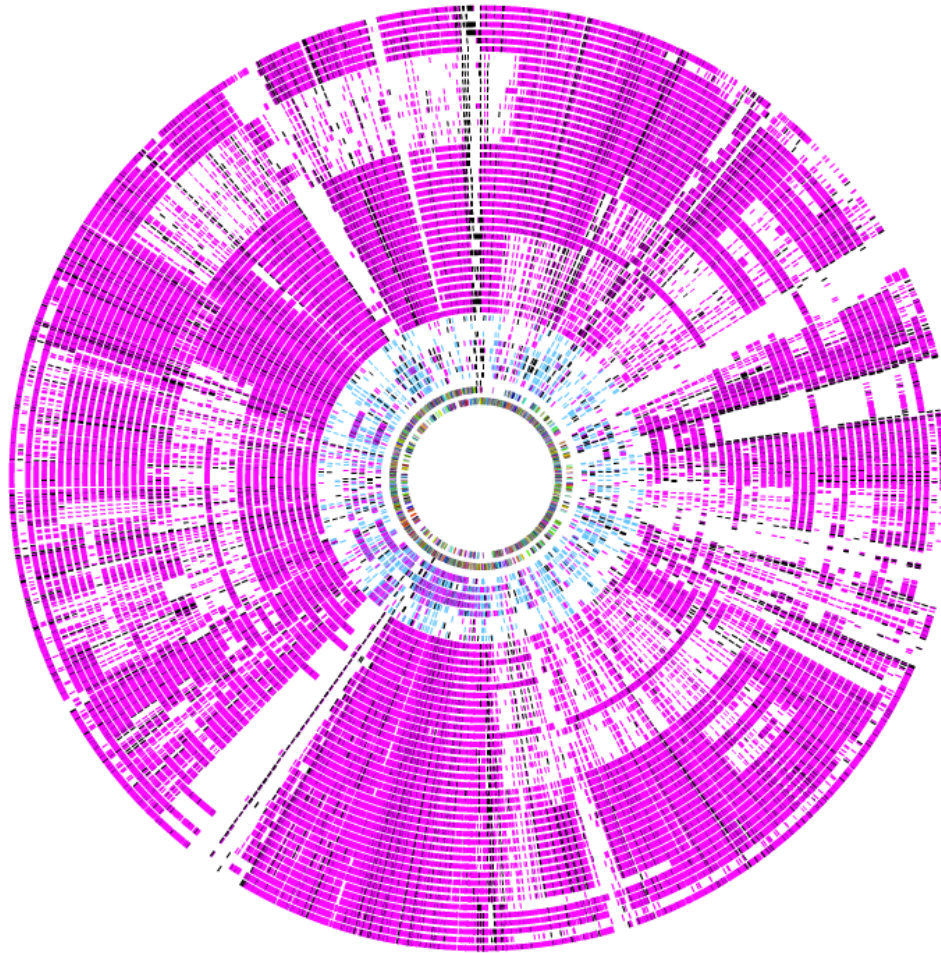
Phylogenetic relationships between the 230 Isolates



Whole genome sequence and MLST

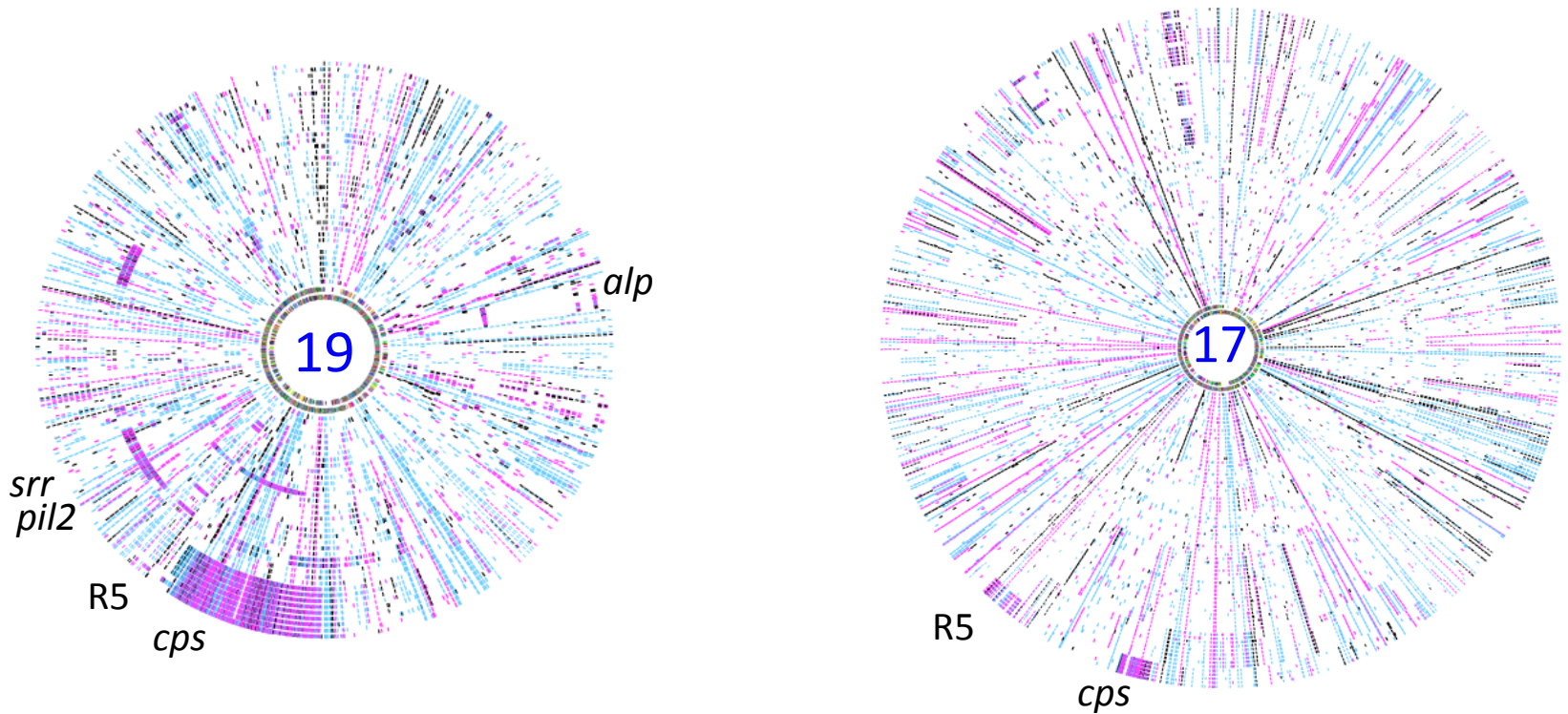


SNP distribution in 63 representative isolates



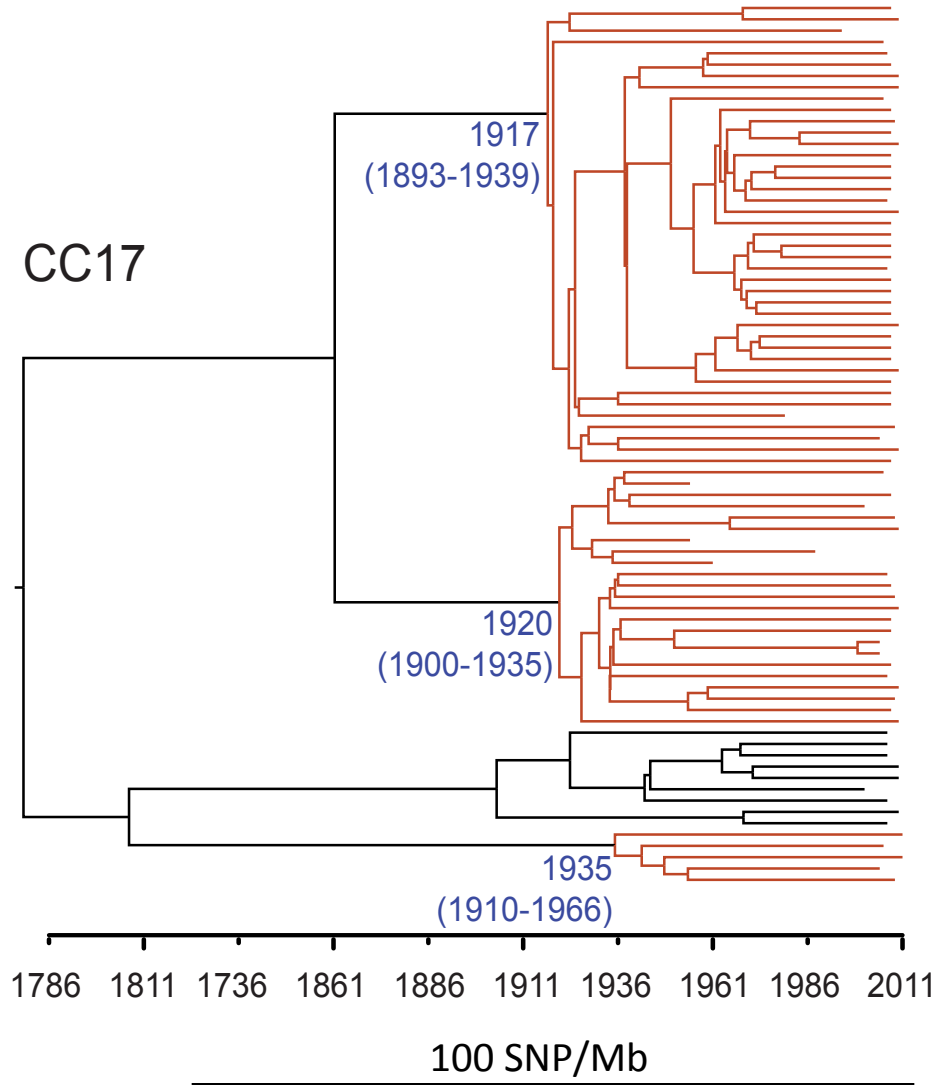
- ⇒ All parts of the genome have recombined
- ⇒ A true phylogeny of the species is not feasible

Recombination within CCs

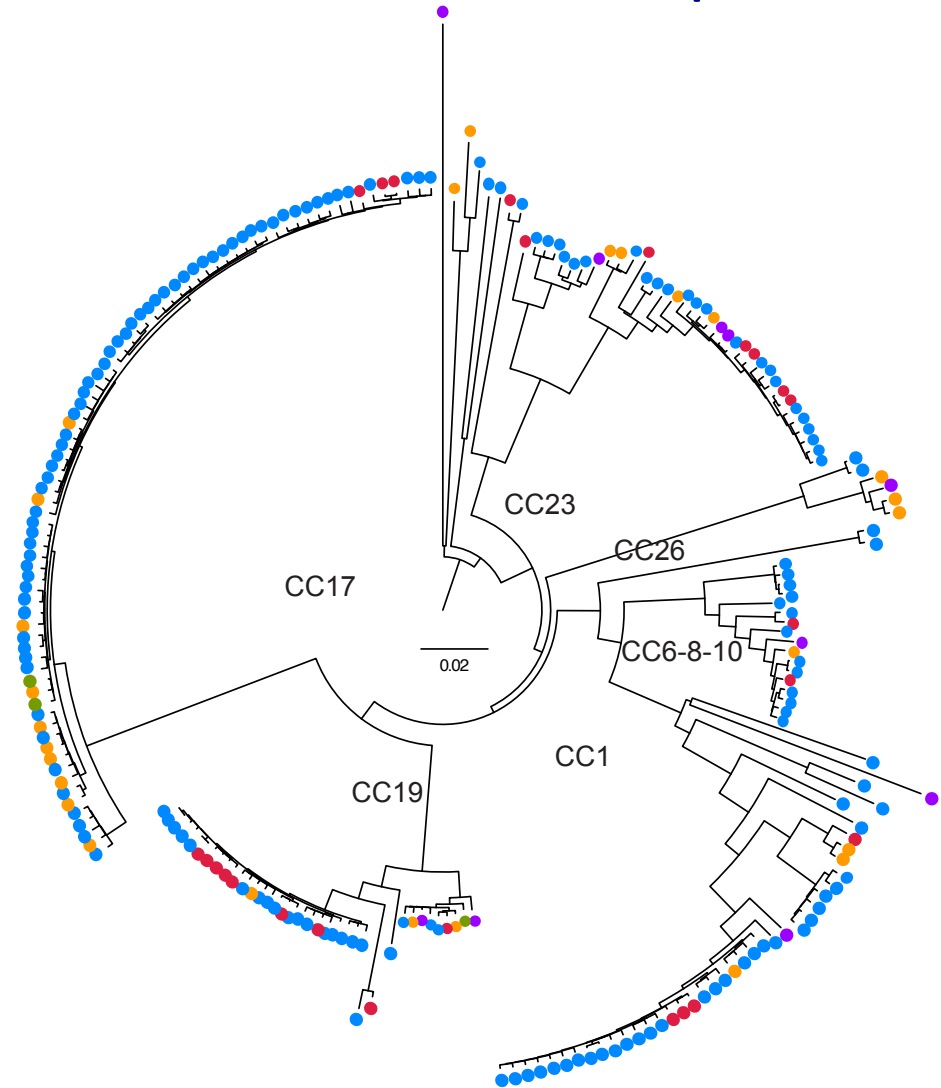


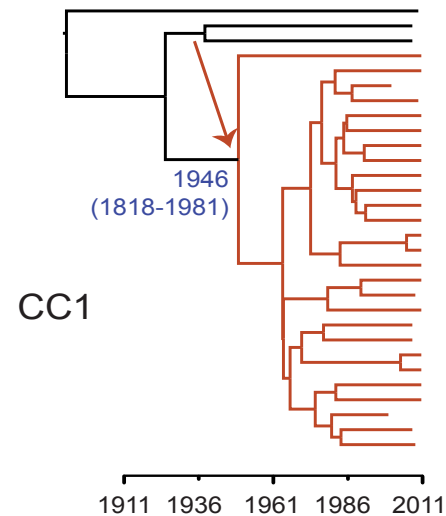
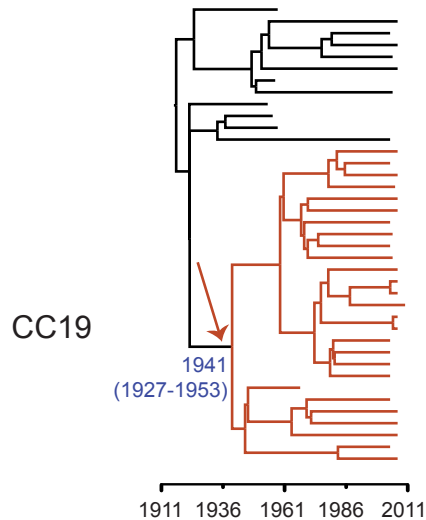
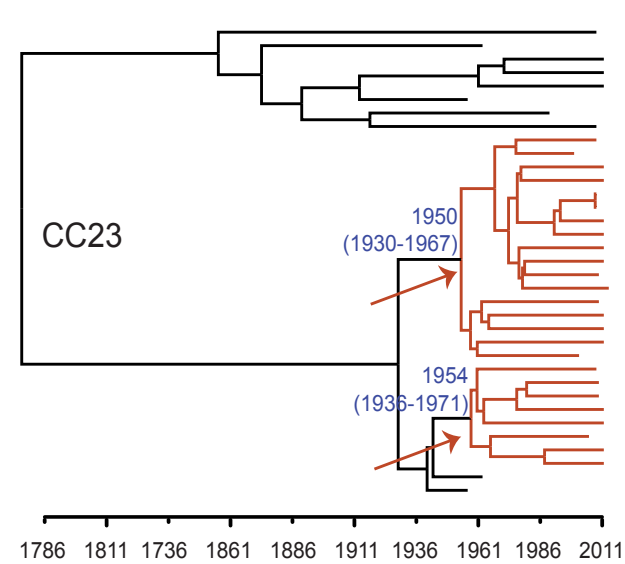
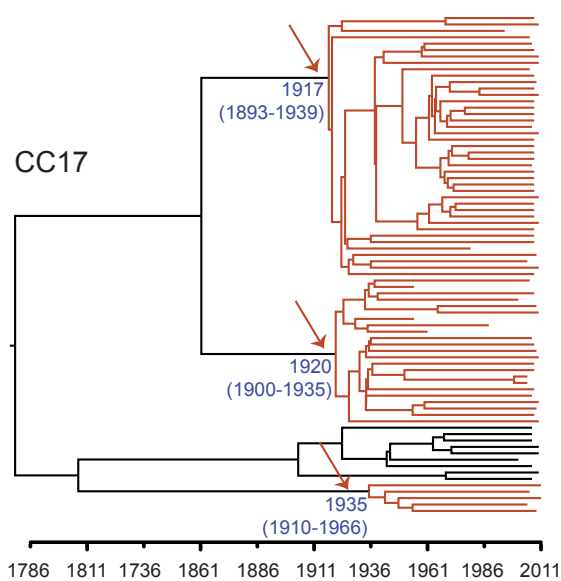
- ⇒ Recombination involves mostly regions encoding antigens
- ⇒ Low level of recombination among the hypervirulent CC17 strains

Time frame for the emergence of clones and complexes



Bayesian analysis





- Similar timeframe for the origin of the 4 major clonal complex
- Timing for the emergence of major clones correspond to the emergence of GBS infections.

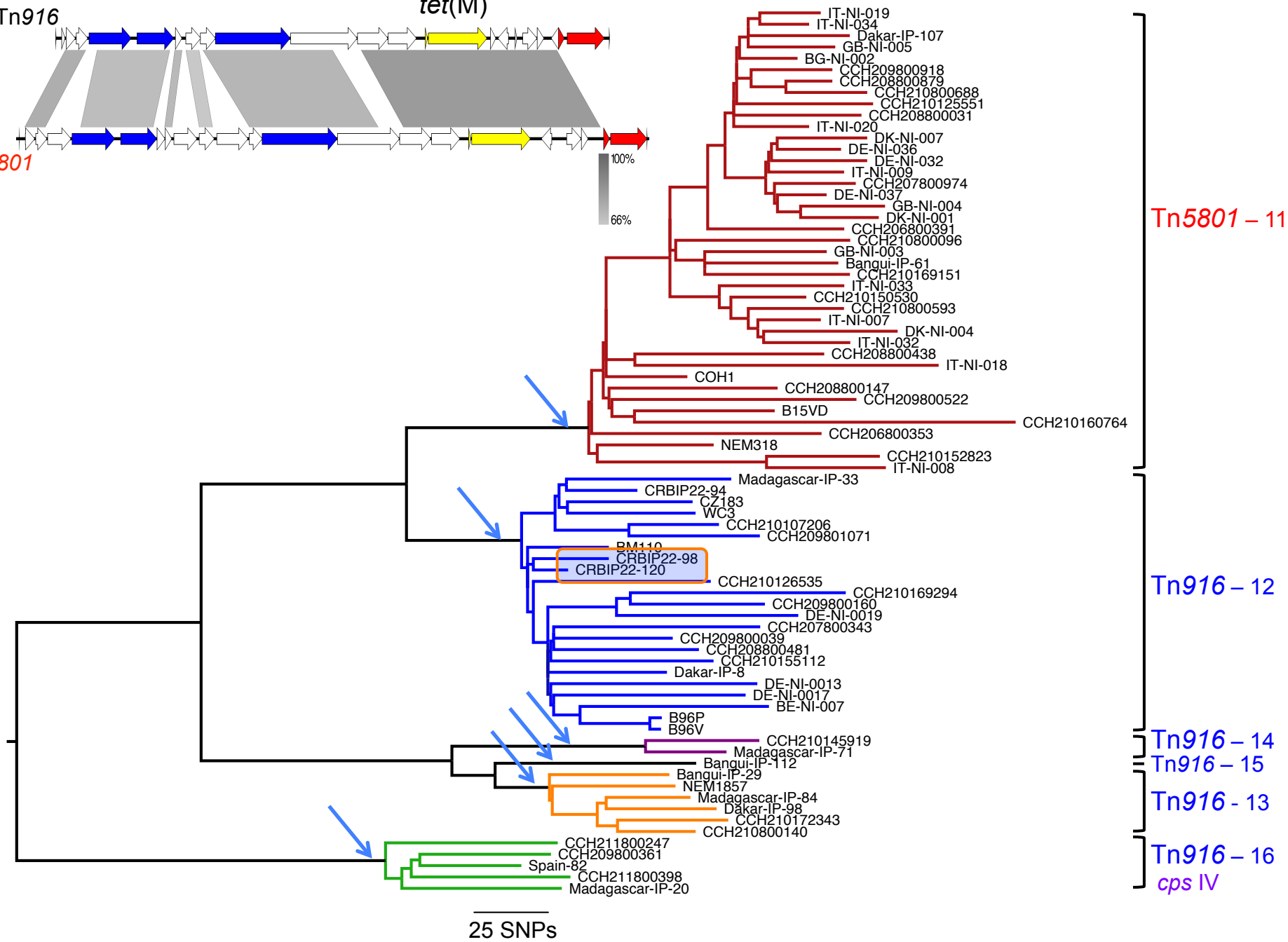
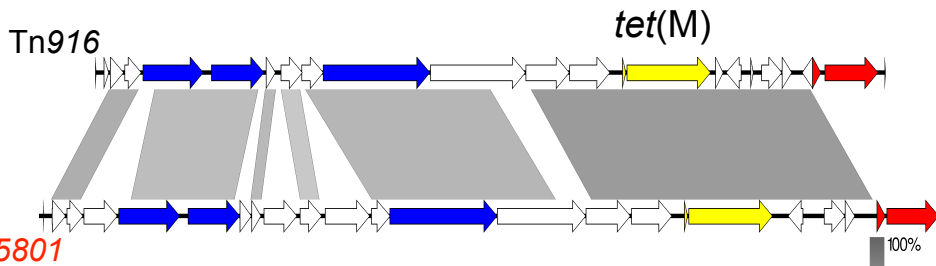
Antibiotic resistance in GBS

In all studies:

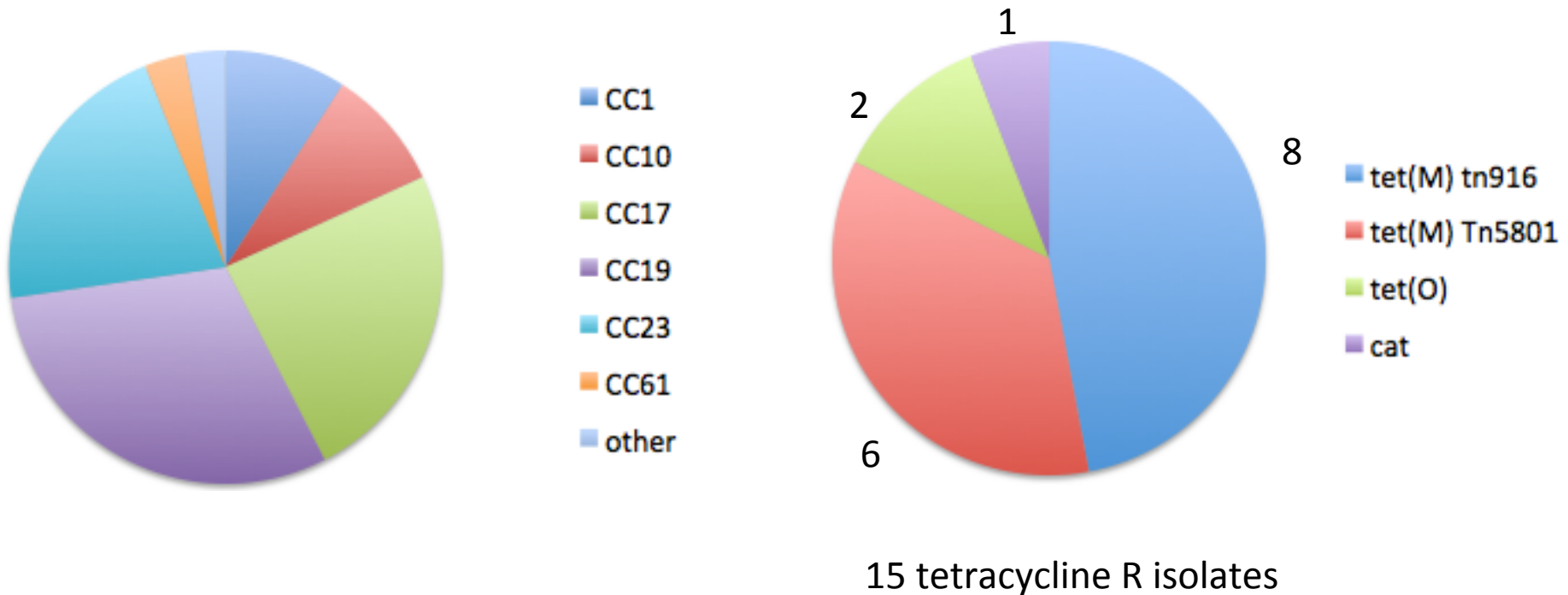
- More than 80 % of human isolates are tetracycline resistant

In the 230 analysed genomes

- 190 isolates express the *tet(M)* gene and 11 the *tet(O)* gene



33 strains from the CIP isolated between 1953 and 1964



- ST distribution is not significantly different from today's distribution
- Tetracycline resistance is present in half the isolates

Few facts

- Tetracycline a broad spectrum antibiotic extensively used starting in the 1950s
- The high rate of tetracycline resistance is specific to human strains compared to bovine strains (but not the use of tetracycline)
- Tetracycline is today rarely used but the tetracycline resistant remains at a high rate.

Proposed scenario for the emergence of GBS neonatal infections

Before 1950: a diverse population of GBS tetracycline sensitive (unknown)

1950: Extensive use of tetracycline

1. Selection of TcR isolates by gain of mobile genetic element
2. Create a niche by eliminating TcS GBS and by altering the gut microbiota
3. Among TcR clones selection of those with higher colonization and dissemination properties
4. Worldwide dissemination of few TcR clones with higher virulence potential

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