

# Be a QPS or not to be, that is the Quirky Paradox of Safety

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# The role of microorganisms in food production

- Fermentation and Preservation

Extended shelf life: by producing compounds that inhibit the growth of spoilage and pathogenic microorganisms

Safety: the acidic environment created by fermentation makes it difficult for harmful pathogens to survive

- Aroma and Texture Enhancement

Flavour profiles: contribute to the development of complex flavors and aromas in products

Texture modification: exopolysaccharides production affecting the texture

- Production of Bioactive Compounds

Vitamins and aminoacids: enhancing the nutritional profile

Antioxidants and antimicrobial compounds: can contribute to overall health and disease prevention

- Nutritional Improvements

Enhanced digestibility: breaks down complex molecules, making nutrients more accessible and easier to digest

Probiotics: fermented products can contain live beneficial bacteria which can improve gut health

- Cultural and Economic Significance

Culinary traditions: fermented foods are integral to many cultural cuisines and traditional diets around the world, contributing to culinary diversity and heritage

Economic value: the fermentation industry is significant economically, with large-scale production of foods and contributing to the global food market

- Sustainability

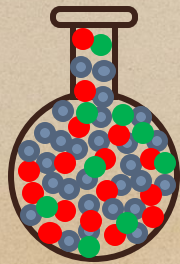
Reduction of food waste: fermentation can help in utilizing surplus produce and reducing food waste

# Biodiversity Loss and Natural Starters

The increased use of stringent hygiene practices in food manufacturing has led to a biodiversity loss in raw matrices and production environments. To counteract this, starter cultures, mainly LAB, are added to the raw material to facilitate and safely carry out the fermentation

## Selected starters

- Made up of two or a few well known selected strains, associated in the laboratory
- Replace the native microbiota with simpler microbiota
- Reduce the microbial biodiversity in products and processing environments causing genetic erosion in the processing environments



## Natural biodiverse starters

- Made up of different microbial species represented by an undefined number of strains (even uncharacterised), in natural balance
- Contribute to the conservation of microbial biodiversity and uniqueness of traditional and PDO products
- Strengthen the link between product and production area (autochthony)



# Starters in PDO productions

According to the PDO (Protected Designation of Origin) regulation, for typical cheesemaking in Sardinia different methods can be used to ensure the authenticity of the product by preserving the traditional practices and specific characteristics of the production area

The PDO regulation allows a relative flexibility:

Pecorino Sardo: use of thermised/pasteurised milk and, if needed, inoculated with selected starters isolated from the production area

Pecorino Romano: use of thermised milk and a natural starter culture (*scotta-innesto*), possibly supplemented with selected starters isolated from the production area

Fiore Sardo: use of only raw milk and, if needed, inoculated with natural starters like *scotta-innesto* or *latto-innesto*



# QPS in the starters

The EFSA introduced, in 2007, the definition of Qualified Presumption of Safety (QPS) for microorganisms that can be intentionally added into the food and feed chain. The QPS list is revised, with an overall assessment carried out by the BIOHAZ Panel, every 3 years and updated every 6 months, and is based on these main points:

Taxonomic identification

Body of knowledge

Safety

The taxonomic units (TUs) of microorganisms that are not well identified, and for which it is not possible to conclude whether they pose a safety concern, are not considered suitable for QPS status and some must undergo a full safety assessment

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**STATEMENT**

 **JOURNAL**

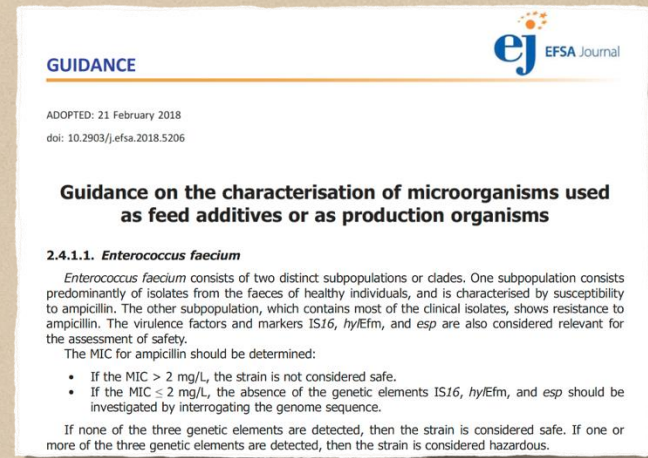
**Update of the list of qualified presumption of safety (QPS)  
recommended microbiological agents intentionally added to  
food or feed as notified to EFSA 20: Suitability of taxonomic  
units notified to EFSA until March 2024**

# Can non-QPS be food starters?

Among the LAB, *Enterococcus faecium* is currently excluded from the QPS list due to its ambiguous hospital (Clade A, isolated from clinical patients) or non-hospital (Clade B, from healthy individuals) origin, and prior to be used as food/feed additive must be tested for safety

A natural starter biodiverse culture (23 strains) obtained from raw ewe milk, without heat treatment or selection of pro-technological microorganisms, included strains belonging to non-QPS species. Following the current EFSA guidelines, the natural starter culture should not be used for food production

- *Enterococcus faecium*
- *Enterococcus faecalis*
- *Enterococcus durans*
- *Lactocaseibacillus paracasei*
- *Streptococcus gallolyticus* subsp. *macedonicus*
- *Streptococcus equinus*
- *Streptococcus lutetiensis*
- *Streptococcus oralis*
- *Streptococcus salivarius*



# Can non-QPS be food starters?

All the *Enterococcus* and *Streptococcus* strains found in the natural starter culture revealed antibiotic sensitivity below the breakpoints indicated by EFSA, EUCAST, and CLSI, and absence of the genetic elements *Stx16*, *hlyEfm*, and *esp*

Bacteria Tested	Culture	Antibiotics Tested								
		Penicillins AMP	Macrolides ERY	Amphenicols CHL	Oxazolidinones LZD	Tetracyclines TET	Glycycyclines TGC	Glycopeptides VAN TEI	Fluoroquinolones CIP	
<i>E. faecium</i>	NSC	S <sup>1</sup>	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	
	LNSC	S <sup>1</sup>	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	
<i>E. faecalis</i>	NSC	S <sup>1</sup>	I <sup>2</sup>	S <sup>2</sup>	S <sup>1</sup>	R <sup>2</sup>	S <sup>1</sup>	S <sup>1</sup>	S <sup>1</sup>	
	LNSC	S <sup>1</sup>	I <sup>2</sup>	S <sup>2</sup>	S <sup>1</sup>	R <sup>2</sup>	S <sup>1</sup>	S <sup>1</sup>	S <sup>1</sup>	
<i>E. durans</i>	NSC	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	
	LNSC	S <sup>1</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>1</sup>	S <sup>2</sup>	S <sup>1</sup>	S <sup>1</sup>	S <sup>1</sup>	

NSC, natural starter culture; LNSC, Lyophilized natural starter culture (LNSC). AMP, Ampicillin; PEN, Penicillin; AZI, Azithromycin; ERY, Erythromycin; FEP, Cefepime; FOT, Cefotaxime; AXO, Ceftriaxone; CHL, Chloramphenicol; CLI, Clindamycin; ETP, Ertapenem; MERO, Meropenem; LEVO, Levofloxacin; LZD, Linezolid; TET, Tetracycline; TGC, Tigecycline; VAN, Vancomycin; TEI, Teicoplanin; SYN, Quinupristin/Dalfopristin; CIP, Ciprofloxacin; S, sensitive; R, resistant; I, intermediate; n.t., not tested; abs., the species was absent in the culture. <sup>1</sup> Breakpoint by EUCAST 2023. <sup>2</sup> Breakpoint by CLSI 2020.

Bacteria Tested	Culture	Antibiotics Tested															
		Penicillins		Macrolides		Cephalosporins			Amphenicols	Lincosamides	Carbapenems		Fluoroquinolones	Oxazolidinones	Tetracyclines	Glycopeptides	Streptogramins
		AMP	PEN	AZI	ERY	FEP	FOT	AXO	CHL	CLI	ETP	MERO	LEVO	LZD	TET	VAN	SYN
<i>S. gallolyticus macedonicus</i>	NSC	n.t.	S <sup>2</sup> *	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
	LNSC	n.t.	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
<i>S. equinus</i>	NSC	n.t.	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
	LNSC	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.
<i>S. lutetiensis</i>	NSC	n.t.	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
	LNSC	n.t.	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
<i>S. oralis</i>	NSC	S <sup>1</sup>	S <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	I <sup>2</sup>
	LNSC	n.t.	S <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	n.t.
<i>S. salivarius</i>	NSC	S <sup>1</sup>	I <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>	S <sup>2</sup>
	LNSC	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.	abs.

NSC, natural starter culture; LNSC, Lyophilized natural starter culture (LNSC). AMP, Ampicillin; PEN, Penicillin; AZI, Azithromycin; ERY, Erythromycin; FEP, Cefepime; FOT, Cefotaxime; AXO, Ceftriaxone; CHL, Chloramphenicol; CLI, Clindamycin; ETP, Ertapenem; MERO, Meropenem; LEVO, Levofloxacin; LZD, Linezolid; TET, Tetracycline; TGC, Tigecycline; VAN, Vancomycin; TEI, Teicoplanin; SYN, Quinupristin/Dalfopristin; CIP, Ciprofloxacin; S, sensitive; R, resistant; I, intermediate; n.t., not tested; abs., the species was absent in the culture. \* one isolate was intermediate. <sup>1</sup> Breakpoint by EUCAST 2023. <sup>2</sup> Breakpoint by CLSI 2020.

# ... and food starters can be non-QPS?

Also other LAB can be used as starters, or can be found, naturally, in the raw matrices, production environments, natural starter, and maybe in products



BIOHAZ statement on QPS: suitability of taxonomic units notified until September 2022

EFSA JOURNAL

## ***Streptococcus oralis***

### *Identity*

*Streptococcus oralis* is a bacterial species with Standing in Nomenclature (Bridge and Sneath, 1982). Based on whole genome sequence analysis three subspecies are recognised: *S. oralis* subsp. *dentisani*, *S. oralis* subsp. *tigurinus* and *S. oralis* subsp. *oralis* (Jensen et al., 2016; Oren and Garrity, 2017).

### *Body of knowledge*

*S. oralis* is part of the normal microbiota of the oropharyngeal, nasal, gastrointestinal and genitourinary tracts and has a 'probiotic' effect providing protection against invading pathogens in the oral cavity (reviewed by Okahashi et al., 2022a; Bidossi et al., 2018). Strains of this species are used as oral 'probiotics', but strains of the same species are responsible for human infections.

### *Safety concerns*

*S. oralis* may cause various diseases such as meningitis, endocarditis and bloodstream infections in which streptococcal surface proteins and other virulence factors might be involved (Basaranoglu et al., 2019; Cruz Cardoso et al., 2021; Nakamura et al., 2021; Okahashi et al., 2022b).

### *Conclusion on a recommendation for QPS status*

*S. oralis* is not recommended for QPS status due to safety concerns.

# The Paradox of non-QPS in natural Consortia

In traditional and PDO productions the challenge faced by artisanal manufacturers is to use their own natural starter cultures to obtain a food with peculiar sensory characteristics and more related to the territory of production (autochthony), also preserving traditional technological methods and microbial diversity

## The Paradox:

The presence of non-QPS microbial species in natural starter cultures obtained from raw matrices, which are inherently part of the traditional production processes, creates a paradox. Artisanal manufacturers may not be allowed to produce or use their own natural starter cultures, as they likely include non-QPS species, but they must comply with the regulations

An answer to this paradox is needed to allow the use of these natural microbial consortia in food production while ensuring food safety



# Addressing the Paradox

The use of microorganisms in food production is a critical aspect of ensuring food safety and quality. The QPS list provides a valuable framework for evaluating the safety of microorganisms intentionally added to food or feed

For some of the species not recommended for QPS status, additional testing is required to ensure the absence of antibiotic resistance genes and virulence factors. While this is a prudent measure to safeguard public health, the lack of specific guidelines for other LAB highlights the need for more comprehensive and flexible regulations

The paradox emerges when the current guidelines prevent the use of these natural starters, as they do not comply with the QPS list requirements. Artisanal manufacturers, who rely on these traditional production methods, may no longer be able to use their own natural starter cultures, jeopardizing the integrity and diversity (also microbial) of their products

Which guidelines should be followed? The PDO regulation mandates the use of traditional techniques (i.e. natural cultures), while EFSA imposes restrictions for food safety reasons. This conflict highlights the challenge of balancing heritage with modern safety standards

# Addressing the Paradox

In conclusion, while the QPS framework serves an important purpose in safeguarding food safety, it should evolve to accommodate the realities of traditional food productions and the inherent value of microbial diversity

By embracing the complexities of natural fermentation processes and the microbial communities involved, a revision of the criteria for the QPS status should be considered in order to find a way for a more inclusive and sustainable food system that honors both safety and tradition

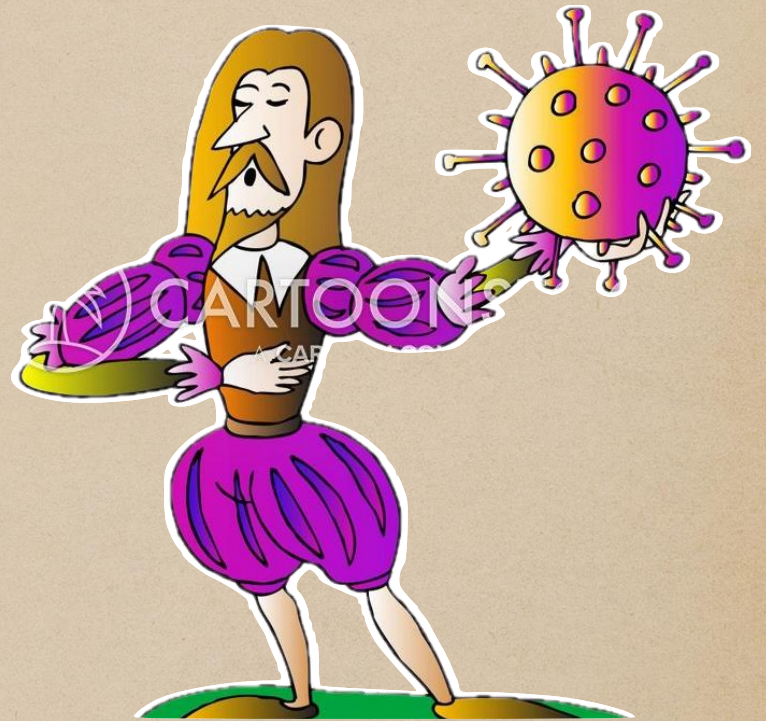


# Thanks for your attention

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